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(54) Title: PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

#### (57) Abstract

By this invention, compositions and methods of use related to  $\beta$ -ketoacyl-ACP synthase of special interest are synthases obtainable from *Cuphea* species. Amino acid and nucleic acid for synthase protein factors are provided, as well as methods to utilize such sequences in constructs for production of genetically engineered plants having altered fatty acid compositions. Of particular interest is the expression of synthase protein factors in conjunction with expression of plant medium-chain acyl-ACP thioesterases for production of increased levels and/or modified ratios of medium-chain fatty acids in oils of transgenic plant seeds.

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# PLANT FATTY ACID SYNTHASES AND USE IN IMPROVED METHODS FOR PRODUCTION OF MEDIUM-CHAIN FATTY ACIDS

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#### INTRODUCTION

#### Field of Invention

The present invention is directed to genes encoding plant fatty acid synthase enzymes relevant to fatty acid synthesis in plants, and to methods of using such genes in combination with genes encoding plant medium-chain preferring thioesterase proteins. Such uses provide a method to increase the levels of medium-chain fatty acids that may be produced in seed oils of transgenic plants.

#### Background

Higher plants synthesize fatty acids via a common metabolic pathway. In developing seeds, where fatty acids attached to triglycerides are stored as a source of energy for further germination, the fatty acid synthesis pathway is located in the plastids. The first step is the formation of acetyl-ACP (acyl carrier protein) from acetyl-CoA and ACP catalyzed by a short chain preferring condensing enzyme, ß-ketoacyl-ACP synthase (KAS) III. Elongation of acetyl-ACP to 16- and 18- carbon fatty acids involves the cyclical action of the following sequence of reactions: condensation with a two-carbon unit from malonyl-ACP to form a longer ß-ketoacyl-ACP (ß-ketoacyl-ACP synthase), reduction of the

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keto-function to an alcohol (ß-ketoacyl-ACP reductase), dehydration to form an enoyl-ACP (ß-hydroxyacyl-ACP dehydrase), and finally reduction of the enoyl-ACP to form the elongated saturated acyl-ACP (enoyl-ACP reductase). ß-ketoacyl-ACP synthase I (KAS I), is primarily responsible for elongation up to palmitoyl-ACP (C16:0), whereas ß-ketoacyl-ACP synthase II (KAS II) is predominantly responsible for the final elongation to stearoyl-ACP (C18:0).

Genes encoding peptide components of ß-ketoacyl-ACP synthases I and II have been cloned from a number of higher plant species, including castor (*Ricinus communis*) and *Brassica* species (USPN 5,510,255). KAS I activity was associated with a single synthase protein factor having an approximate molecular weight of 50 kD (synthase factor B) and KAS II activity was associated with a combination of two synthase protein factors, the 50 kD synthase factor B and a 46 kd protein designated synthase factor A. Cloning and sequence of a plant gene encoding a KAS III protein has been reported by Tai and Jaworski (*Plant Physiol*. (1993) 103:1361-1367).

The end products of plant fatty acid synthetase activities are usually 16- and 18-carbon fatty acids. There are, however, several plant families that store large amounts of 8- to 14-carbon (medium-chain) fatty acids in their oilseeds. Recent studies with Umbellularia californica (California bay), a plant that produces seed oil rich in lauric acid (12:0), have demonstrated the existence of a medium-chain-specific isozyme of acyl-ACP thioesterase

in the seed plastids. Subsequent purification of the 12:0-ACP thioesterase from Umbellularia californica led to the cloning of a thioesterase cDNA which was expressed in seeds of Arabidopsis and Brassica resulting in a substantial accumulation of lauric acid in the triglyceride pools of these transgenic seeds (USPN 5,512,482). These results and subsequent studies with medium-chain thioesterases from other plant species have confirmed the chain-length-determining role of acyl-ACP thioesterases during de novo fatty acid biosynthesis (T. Voelker (1996) Genetic Engineering, Ed. J. K. Setlow, Vol. 18, pgs. 111-133).

#### DESCRIPTION OF THE FIGURES

Figure 1. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-2 are provided. 15 Figure 2. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor B clone chKAS B-31-7 are provided. Figure 3. DNA and translated amino acid sequence of Cuphea hookeriana KAS factor A clone chKAS A-2-7 are provided. Figure 4. DNA and translated amino acid sequence of Cuphea 20 hookeriana KAS factor A clone chKAS A-1-6 are provided. Figure 5. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/7-8 are provided. Figure 6. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor B clone cpuKAS B/8-7A are provided. Figure 7. DNA and translated amino acid sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p7-6A are provided. Figure 8. Preliminary DNA sequence of Cuphea pullcherrima KAS factor A clone cpuKAS A/p8-9A is provided.

Figure 9. DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 are provided.

- Figure 10. The activity profile for purified cpuKAS B/8-7A using various acyl-ACP substrates is provided.
- 5 Figure 11. The activity profile for purified chKAS A-2-7 and chKAS A-1-6 using various acyl-ACP substrates is provided.
  - Figure 12. The activity profile for purified castor KAS factor A using various acyl-ACP substrates is provided.
- factor B using various acyl-ACP substrates is provided.

  Figure 14. A graph showing the number of plants arranged according to C8:0 content for transgenic plants containing CpFatBl versus transgenic plants containing CpFatBl versus transgenic plants containing CpFatBl + chKAS
- 15 A-2-7 is provided.

  Figure 15. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS)

A-2-7 plants) are provided.

- Figure 16. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5401-9 (chKAS A-2-7 plants) are provided.
  - Figure 17. Graphs showing the %C10/%C8 ratios in transgenic plants containing ChFatB2 (4804-22-357) and in plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.
    - Figure 18. Graphs showing the %C10 + %C8 contents in transgenic plants containing ChFatB2 (4804-22-357) and in

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plants resulting from crosses between 4804-22-357 and 5413-17 (chKAS A-2-7 + CpFatB1 plants) are provided.

Figure 19. Graphs showing the %C12:0 in transgenic plants containing Uc FatBl (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 20. Graph showing the relative proportions of C12:0 and C14:0 fatty acids in the seeds of transgenic plants containing Uc FatB1 (LA86DH186) and in plants resulting from crosses with wild type (X WT) and with lines expressing Ch KAS A-2-7.

Figure 21. Graphs showing the %C18:0 in transgenic plants containing Garm FatB1 (5266) and in seeds of plants resulting from crosses with wild type (X WT) and with lines expressing

15 Ch KAS A-2-7.

Figure 22. The activity profile of Ch KAS A in protein extracts from transgenic plants containing Ch KAS A-2-7. Extracts were preptreated with the indicated concentrations of cerulenin.

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#### SUMMARY OF THE INVENTION

By this invention, compositions and methods of use related to ß-ketoacyl-ACP synthase (KAS) are provided. Also of interest are methods and compositions of amino acid and nucleic acid sequences related to biologically active plant synthase(s).

In particular, genes encoding KAS protein factors A and B from *Cuphea* species are provided. The KAS genes are of interest for use in a variety of applications, and may be

used to provide synthase I and/or synthase II activities in transformed host cells, including bacterial cells, such as E. coli, and plant cells. Synthase activities are distinguished by the preferential activity towards longer and shorter acyl-ACPs as well as by the sensitivity towards the KAS specific inhibitor, cerulenin. Synthase protein preparations having preferential activity towards medium chain length acyl-ACPs are synthase I-type or KAS I. The KAS I class is sensitive to inhibition by cerulenin at concentrations as low as 1 µM. Synthases having preferential activity towards longer chain length acyl-ACPs are synthase II-type or KAS II. The KAS enzymes of the II-type are also sensitive to cerulenin, but at higher concentrations (50μM). Synthase III-type enzymes have preferential activity towards short chain length acyl-ACPs and are insensitive to cerulenin inhibition.

Nucleic acid sequences encoding a synthase protein may be employed in nucleic acid constructs to modulate the amount of synthase activity present in the host cell, especially the relative amounts of synthase I-type, synthase II-type and synthase III-type activity when the host cell is a plant host cell. In addition, nucleic acid constructs may be designed to decrease expression of endogenous synthase in a plant cell as well. One example is the use of an antisense synthase sequence under the control of a promoter capable of expression in at least those plant cells which normally produce the enzyme.

Of particular interest in the present invention is the coordinate expression of a synthase protein with the

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expression of thioesterase proteins. For example, coordinated expression of synthase factor A and a medium-chain thioesterase provides a method for increasing the level of medium-chain fatty acids which may be harvested from transgenic plant seeds. Furthermore, coordinated expression of a synthase factor A gene with plant medium-chain thioesterase proteins also provides a method by which the ratios of various medium-chain fatty acids produced in a transgenic plant may be modified. For example, by expression of a synthase factor A, it is possible to increase the ratio of C10/C8 fatty acids which are produced in plant seed oils as the result of expression of a thioesterase having activity on C8 and C10 fatty acids.

## DETAILED DESCRIPTION OF THE INVENTION

A plant synthase factor protein of this invention includes a sequence of amino acids or polypeptide which is required for catalyzation of a condensation reaction between an acyl-ACP having a chain length of C2 to C16 and malonyl-ACP in a plant host cell. A particular plant synthase factor protein may be capable of catalyzing a synthase reaction in a plant host cell (for example as a monomer or homodimer) or may be one component of a multiple peptide enzyme which is capable of catalyzing a synthase reaction in a plant host cell, i.e. one peptide of a heterodimer.

Synthase I (KAS I) demonstrates preferential activity towards acyl-ACPs having shorter carbon chains, C2-C14 and is sensitive to inhibition by cerulenin at concentrations of 1µM. Synthase II (KAS II) demonstrates preferential

activity towards acyl-ACPs having longer carbon chains, C14-C16, and is inhibited by concentrations of cerulenin (50µM). Synthase III demonstrates preferential activity towards acyl-CoAs having very short carbon chains, C2 to C6, and is insensitive to inhibition by cerulenin.

Synthase factors A and B, and synthase III proteins obtained from medium-chain fatty acid producing plant species of the genus Cuphea are described herein. As described in the following Examples, synthase A from C. hookeriana is naturally expressed at a high level and only in the seeds. C. hookeriana synthase B is expressed at low levels in all tissues examined. Expression of synthase A and synthase B factors in E. coli and purification of the resulting proteins is employed to determine activity of the various synthase factors. Results of these analyses indicate that synthase factor A from Cuphea hookeriana has the greatest activity on 6:0-ACP substrates, whereas synthase factor B from Cuphea pullcherrima has greatest activity on 14:0-ACP. Similar studies with synthase factors A and B from castor demonstrate similar activity profiles 20 between the factor B synthase proteins from Cuphea and castor. The synthase A clone from castor, however, demonstrates a preference for 14:0-ACP substrate.

Expression of a Cuphea hookeriana KAS A protein in transgenic plant seeds which normally do not produce medium-chain fatty acids does not result in any detectable modification of the fatty acid types and contents produced in such seeds. However, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a

medium-chain acyl-ACP thioesterase capable of providing for production of C8 and C10 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are observed. In addition, where significant amounts of C8 and C10 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C8 and C10 fatty acids that are obtained. For example, an increased proportion of C10 fatty acids may be obtained by co-expression of Cuphea hookeriana ChFatB2 thioesterase and a chKAS A synthase factor proteins.

Furthermore, when Cuphea hookeriana KAS A protein is expressed in conjunction with expression of a medium-chain acyl-ACP thioesterase capable of providing for production of C12 fatty acids in plant seed oils, increases in the levels of medium-chain fatty acids over the levels obtainable by expression of the medium-chain thioesterase alone are also observed. In addition, where significant amounts of C12 and C14 fatty acids are produced as the result of medium-chain thioesterase expression, co-expression of a Cuphea KAS A protein also results in an alteration of the proportion of the C12 and C14 fatty acids that are obtained. For example, an increased proportion of C12 fatty acids may be obtained by co-expression of Uc FatB1 thioesterase and a chKAS A synthase factor proteins.

However, when Cuphea hookeriana KAS A protein is expressed in conjunction with the expression of a long-chain

acyl-ACP thioesterase capable of providing for production of C18 and C18:1 fatty acids in plant seed oils, no effect on the production of long chain fatty acids was observed.

Furthermore, when plants transformed to express a long chain acyl-ACP thioesterase from mangosteen (GarmFatA1, U.S. Patent Application No. 08/440,845), which preferentially hydrolyzes C18:0 and C18:1 fatty acyl-ACPs, are crossed with nontransformed control plants, a significant reduction in the levels of C18:0 is obtained. Similar reductions are also observed in the levels of C18:0 in the seeds of plants resulting from crosses between plants transformed to express the GarmFatA1 and plants expressing the Cuphea hookeriana KAS A protein.

Thus, the instant invention provides methods of increasing and/or altering the medium-chain fatty acid compositions in transgenic plant seed oils by co-expression of medium-chain acyl-ACP thioesterases with synthase factor proteins. Furthermore, various combinations of synthase factors and medium-chain thioesterases may be achieved depending upon the particular fatty acids desired. For example, for increased production of C14 fatty acids, synthase protein factors may be expressed in combination with a C14 thioesterase, for example from Cuphea palustris or nutmeg may be employed (WO 96/23892). In addition, thioesterase expression may be combined with a number of different synthase factor proteins for additional effects on medium-chain fatty acid composition.

Synthases of use in the present invention include modified amino acid sequences, such as sequences which have

been mutated, truncated, increased and the like, as well as such sequences which are partially or wholly artificially synthesized. The synthase protein encoding sequences provided herein may be employed in probes for further screening or used in genetic engineering constructs for transcription or transcription and translation in host cells, especially plant host cells. One skilled in the art will readily recognize that antibody preparations, nucleic acid probes (DNA and RNA) and the like may be prepared and 10 used to screen and recover synthases and/or synthase nucleic acid sequences from other sources. Typically, a homologously related nucleic acid sequence will show at least about 60% homology, and more preferably at least about 70% homology, between the R. communis synthase and the given plant synthase of interest, excluding any deletions which may be present. Homology is determined upon comparison of sequence information, nucleic acid or amino acid, or through hybridization reactions.

Recombinant constructs containing a nucleic acid sequence encoding a synthase protein factor or nucleic acid sequences encoding a synthase protein factor and a medium-chain acyl-ACP thioesterase may be prepared by methods well known in the art. Constructs may be designed to produce synthase in either prokaryotic or eukaryotic cells. The increased expression of a synthase in a plant cell, particularly in conjunction with expression of medium-chain thioesterases, or decreasing the amount of endogenous synthase observed in plant cells are of special interest.

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Synthase protein factors may be used, alone or in combination, to catalyze the elongating condensation reactions of fatty acid synthesis depending upon the desired result. For example, rate influencing synthase activity may reside in synthase I-type, synthase II-type, synthase III-type or in a combination of these enzymes. Furthermore, synthase activities may rely on a combination of the various synthase factors described herein.

Constructs which contain elements to provide the transcription and translation of a nucleic acid sequence of interest in a host cell are "expression cassettes".

Depending upon the host, the regulatory regions will vary, including regions from structural genes from viruses, plasmid or chromosomal genes, or the like. For expression in prokaryotic or eukaryotic microorganisms, particularly unicellular hosts, a wide variety of constitutive or regulatable promoters may be employed. Among transcriptional initiation regions which have been described are regions from bacterial and yeast hosts, such as E. coli, B. subtilis, Saccharomyces cerevisiae, including genes such as ß-galactosidase, T7 polymerase, trp-lac (tac), trp E and the like.

An expression cassette for expression of synthase in a plant cell will include, in the 5' to 3' direction of transcription, a transcription and translation initiation control regulatory region (also known as a "promoter") functional in a plant cell, a nucleic acid sequence encoding a synthase, and a transcription termination region.

Numerous transcription initiation regions are available

which provide for a wide variety of constitutive or regulatable, e.g., inducible, transcription of the desaturase structural gene. Among transcriptional initiation regions used for plants are such regions associated with cauliflower mosaic viruses (35S, 19S), and structural genes such as for nopaline synthase or mannopine synthase or napin and ACP promoters, etc. The transcription/ translation initiation regions corresponding to such structural genes are found immediately 5' upstream to the respective start codons. Thus, depending upon the intended use, different promoters may be desired.

Of special interest in this invention are the use of promoters which are capable of preferentially expressing the synthase in seed tissue, in particular, at early stages of seed oil formation. Examples of such seed-specific promoters include the region immediately 5' upstream of a napin or seed ACP genes such as described in USPN 5,420,034, desaturase genes such as described in Thompson et al (Proc. Nat. Acad. Sci. (1991) 88:2578-2582), or a Bce-4 gene such as described in USPN 5,530,194. Alternatively, the use of the 5' regulatory region associated with the plant synthase structural gene, i.e., the region immediately 5' upstream to a plant synthase structural gene and/or the transcription termination regions found immediately 3' downstream to the plant synthase structural gene, may often be desired. general, promoters will be selected based upon their expression profile which may change given the particular application.

In addition, one may choose to provide for the transcription or transcription and translation of one or more other sequences of interest in concert with the expression or anti-sense of the synthase sequence, particularly medium-chain plant thioesterases such as described in USPN 5,512,482, to affect alterations in the amounts and/or composition of plant oils.

When one wishes to provide a plant transformed for the combined effect of more than one nucleic acid sequence of interest, a separate nucleic acid construct may be provided for each or the constructs may both be present on the same plant transformation construct. The constructs may be introduced into the host cells by the same or different methods, including the introduction of such a trait by crossing transgenic plants via traditional plant breeding methods, so long as the resulting product is a plant having both characteristics integrated into its genome.

Normally, included with the DNA construct will be a structural gene having the necessary regulatory regions for expression in a host and providing for selection of transformed cells. The gene may provide for resistance to a cytotoxic agent, e.g. antibiotic, heavy metal, toxin, etc., complementation providing prototrophy to an auxotrophic host, viral immunity or the like. Depending upon the number of different host species into which the expression construct or components thereof are introduced, one or more markers may be employed, where different conditions for selection are used for the different hosts.

The manner in which the DNA construct is introduced into the plant host is not critical to this invention. Any method which provides for efficient transformation may be employed. Various methods for plant cell transformation include the use of Ti- or Ri-plasmids, microinjection, electroporation, liposome fusion, DNA bombardment or the like. In many instances, it will be desirable to have the construct bordered on one or both sides by T-DNA, particularly having the left and right borders, more particularly the right border. This is particularly useful when the construct uses A. tumefaciens or A. rhizogenes as a mode for transformation, although the T-DNA borders may find use with other modes of transformation.

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The expression constructs may be employed with a wide variety of plant life, particularly plant life involved in the production of vegetable oils. These plants include, but are not limited to rapeseed, peanut, sunflower, safflower, cotton, soybean, corn and oilseed palm.

For transformation of plant cells using Agrobacterium, explants may be combined and incubated with the transformed Agrobacterium for sufficient time for transformation, the bacteria killed, and the plant cells cultured in an appropriate selective medium. Once callus forms, shoot formation can be encouraged by employing the appropriate plant hormones in accordance with known methods and the shoots transferred to rooting medium for regeneration of plants. The plants may then be grown to seed and the seed used to establish repetitive generations and for isolation of vegetable oils.

The invention now being generally described, it will be more readily understood by reference to the following examples which are included for purposes of illustration only and are not intended to limit the present invention.

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#### **EXAMPLES**

# Example 1 Cuphea KAS Factor A and B Gene Cloning

Total RNA isolated from developing seeds of Cuphea hookeriana and Cuphea pullcherrima was used for cDNA synthesis in commercial 1-based cloning vectors. cloning each type of KAS gene, approximately 400,000-500,000 unamplified recombinant phage were plated and the plaques transferred to nitrocellulose. For KAS factor B cloning from C. hookeriana, a mixed probe containing Brassica napus KAS factor B and Ricinus communis (Castor) KAS factor B radiolabeled cDNA's was used. Similarly, a mixed probe containing Brassica napus KAS factor A and Ricinus communis KAS factor A cDNA clones was used to obtain C. hookeriana KAS factor A genes. For KASIII, a spinach KASIII cDNA clone obtained from Dr. Jan Jaworski was radiolabeled and used as a probe to isolate a KASIII clone from C. hookeriana. For KAS B and KAS A cloning from C. pullcherrima, C. hookeriana KAS B and KAS A genes chKAS B-2 and chKAS A-2-7 (see below) were radiolabeled and used as 25 probes.

DNA sequence and translated amino acid sequence for Cuphea KAS clones are provided in Figures 1-9. Cuphea hookeriana KAS factor B clones chKAS B-2 and chKAS B-31-7

are provided in Figures 1 and 2. Neither of the clones is full length. Cuphea hookeriana KAS Factor A clones chKAS A-2-7 and chKAS A-1-6 are provided in Figures 3 and 4. chKAS A-2-7 contains the entire encoding sequence for the KAS factor protein. Based on comparison with other plant synthase proteins, the transit peptide is believed to be represented in the amino acids encoded by nucleotides 125-466. chKAS A-1-6 is not a full length clone although some transit peptide encoding sequence is present. Nucleotides 1-180 represent transit peptide encoding sequence, and the mature protein encoding sequence is believed to begin at nucleotide 181.

Cuphea pullcherrima KAS factor B clones cpuKAS B/7-8 and cpuKAS B/8-7A are provided in Figures 5 and 6. Both of the clones contain the entire encoding sequences for the KAS factor B proteins. The first 35 amino acids of cpuKAS B/7-8 are believed to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide 233. The first 39 amino acids of cpuKAS B/8-7A are believed to represent the transit peptide, with the mature protein encoding sequence beginning at nucleotide 209. Cuphea pullcherrima KAS factor A clones cpuKAS A/p7-6A and cpuKAS A-p8-9A are provided in Figures 7 and 8. Both of the clones contain the entire encoding sequences for the KAS factor A proteins. Translated amino acid sequence of cpuKAS A/p7-6A is provided. The mature protein is believed to begin at the lysine residue encoded 595-597, and the first 126 amino acids are believed to represent the transit peptide. DNA sequence of KAS A clone cpuKAS A-p8-9A is preliminary.

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DNA and translated amino acid sequence of Cuphea hookeriana KASIII clone chKASIII-27 is provided in Figure 9. The encoding sequence from nucleotides 37-144 of chKASIII-27 are believed to encode a transit peptide, and the presumed mature protein encoding sequence is from nucleotides 145-1233.

Deduced amino acid sequence of the C. hookeriana KAS factor B and KAS factor A cDNA's reveals strong homology to the Brassica napus and Ricinus communis clones previously reported. The C. hookeriana KAS factor B clone is more homologous to the Ricinus and Brassica KAS factor B clones (94% and 91% respectively) than it is to the Ricinus and Brassica KAS factor A clones (60% for both). Furthermore, the C. hookeriana KAS factor A clone is more homologous to the Ricinus and Brassica KAS factor A clones (85% and 82% respectively) than it is the Ricinus and Brassica KAS factor B clone (60% for both). The C. hookeriana KAS factor B cDNAs designated as chKAS B-2 and chKAS B-31-7 are 96% identical within the mature portion of the polypeptide. Similarly, the deduced amino acid sequence of the mature protein regions of the C. hookeriana KAS factor A clones chKAS A-2-7 and chKAS A-1-6 are 96% identical. The C.pullcherrima KAS clones also demonstrate homology to the R. communis and Brassica napus KAS clones. The mature protein portion of all of the KAS factor A family members in the different Cuphea species are 95% identical. Similarly the

mature protein portion of the KAS factor B genes in Cuphea are also 95-97% identical with each other. However there is only approximately 60% sequence identity between KAS factor B and KAS factor A clones either within the same or different species of Cuphea.

### Example 2 Levels and Patterns of Expression

To examine tissue specificity of KAS expression in Cuphea hookeriana, Northern blot analysis was conducted using total RNA isolated from seed, root, leaf and flower tissue. Two separate but identical blots were hybridized with either chKAS B-31-7 or chKAS A-2-7 coding region probes. The data from this RNA blot analysis indicate that KAS B is expressed at a similar level in all tissues examined, whereas KAS A expression is detected only in the seed. These results also demonstrate a different level of expression for each of the synthases. KAS A is an abundant message, whereas KAS B is expressed at low levels. Furthermore, even under highly stringent hybridization conditions (65\_C, 0.1 X SSC, 0.5% SDS), the KAS A probe hybridizes equally well with two seed transcripts of 2.3 and 1.9 kb. The larger hybridizing band is likely the transcript of the KAS A-2-7 gene since the size of its cDNA is 2046bp, and the number of clones obtained from cDNA screening corresponds well with the apparent mobility of the mRNA and its abundance on the blot.

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# Example 3 Expression of Plant KAS Genes in E.coli

DNA fragments encoding the mature polypeptide of the Cuphea hookeriana KAS A cDNAs and the Cuphea pullcherrima 5 KAS B cDNAs were obtained by PCR and cloned into a QIAexpress expression vector (Qiagene). Experimental conditions for maximum level of expression were determined for all of these clones and the parameters for highest level of soluble fraction were identified. Cells are grown in 10 ECLB media containing 1M sorbitol and 2.5 mM betaine overnight and subcultured as a 1:4 dilution in the same medium. Cells are then grown for 2 hours (to approximately .6-.8 O.D.) and induced with 0.4 mM IPTG and allowed to grow for 5 more hours.

Enzyme activity of the affinity purified recombinant enzymes obtained from over-expression of the chKAS A-2-7 and cpuKAS B/8-7A clones was measured using a wide range of acyl-ACP substrates (6:0- to 16:1-ACP). The activity profile for cpuKAS B/8-7A is provided in Fig.10. The data demonstrate that the enzyme is active with all acyl-ACP substrates examined, although activity on 6:0 to 14:0-ACP substrates is substantially greater than the activity on 16:0 and 16:1 substrates.

The activity profile of the C. hookeriana KAS A clones chKAS A-2-7 and chKAS A-1-6 is provided in Figure 11. The C. 25 hookeriana KAS A clones are most active with C:6, and have the least activity with C:16:0 substrates. However, the activity of this clone on even the preferred C6:0 substrate

is 50 fold lower than the activity of the *C. pullcherrima*KAS B clones.

A fragment containing the mature protein encoding portion of a R. communis KAS factor A clone was also cloned into a QIAexpress expression vector, expressed in E. coli and the enzyme affinity purified as described above. The activity profile for castor KAS A is provided in Figure 12. Highest activity is observed with C14:0 substrates, although some activity is also seen with C6:0 and C16:1. 10 comparison, the activity profile obtained from purified R. communis KAS factor B also using the QIAexpress expression system is provided in Figure 13. The KAS B clone demonstrates substantially higher levels of activity (10 fold and higher) than the R. communis KAS A clone. preference of the KAS factor B for 6:0- to 14:0-ACP substrates is consistent with the previous observations that this protein provides KAS I activity.

## Example 4 KAS and TE Expression in Transgenic Seed

Dehesh et al. (1996) Plant Physiol. 110:203-210) and the chKAS A-2-7 were PCR amplified, sequenced, and cloned into a napin expression cassette. The napin/cp FatB1 and the napin/KAS A-2-7 fusions were ligated separately into the binary vector pCGN1558 (McBride and Summerfelt (Pl.Mol.Biol. (1990) 14:269-276) and transformed into A. tumefaciens, EHA101. The resulting CpFatB1 binary construct is pCGN5400 and the chKAS A-2-7 construct is pCGN5401. Agrobacterium mediated transformation of a Brassica napus canola variety

was carried out as described by Radke et al. (Theor. Appl. Genet. (1988) 75:685-694; Plant Cell Reports (1992) 11:499-505). Several transgenic events were produced for each of the pCGN5400 and pCGN5401 constructs.

A double gene construct containing a napin/cpFatB1 expression construct in combination with a napin/chKAS A-2-7 expression construct was also assembled, ligated into a binary vector and used for co-cultivation of a canola Brassica variety. The binary construct containing the chFatB1 and chKAS A-2-7 expression constructs is pCGN5413.

Fatty acid analysis of 26 transgenic lines containing chKAS A-2-7 (5401 lines) showed no significant changes in the oil content or profile as compared to similar analyses of wild type canola seeds of the transformed variety.

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ratty acid analysis of 36 transgenic lines containing cpFatB1 (5400 lines) showed increased levels of C:8 and C:10 in transgenic seeds. The highest level of C:8 observed in a pool seed sample was 4.2 mol%. The C:10 levels were between 30 and 35% of the C:8 content. Fatty acid analysis of 25 transgenic lines containing the TE/KAS A tandem (5413 lines) demonstrated an overall increase in both C:8 and C:10 levels relative to those observed with TE containing lines (5400) alone. In lines containing the cpFatB1 construct alone, the average level of C:8 average were 1.5 mol%, whereas the C:8 average levels in TE/KAS A tandem containing lines was 2.37 mol%. The ratio of C:8 to C:10 remained constant in both populations. The number of transgenic events relative to the C:8 content are presented in Figure 14. These data show that the transgenic events with tandem TE/KAS A construct

yield more lines with higher levels of C:8 than those events with single TE construct. For example, several lines containing nearly 7 mole% C8 were obtained with the TE/KAS A pCGN5413 construct, whereas the highest C8 containing line from the pCGN5400 TE alone transformation contained 4.2 mole% C8.

Half seed analysis of the T3 generation of transgenic canola plants expressing a ChFatB2 (*C. hookeriana* thioesterase; Dehesh et al. (1996) The Plant Journal 9:167-172) indicate that these plant can accumulate up to 22 weight% (33 mol%) of 8:0 and 10:0 fatty acids (4804-22-357). Segregation analysis shows that these transformants contain two loci and that they are now homozygous. Selected plants grown from these half seeds were transferred into the greenhouse and later crossed with T1 transformants that had been transformed with either Cuphea hookeriana KAS A (5401) alone or KAS A/CpFatB1 double constructs (5413).

Fatty acid analysis of several events resulting from the crosses between transgenic lines containing ChFatB2 (4804-22-357) and chKAS A-2-7 (5401-9), reveal an increase in the ratio of C:10/C:8 levels (Figure 15). This C:10/C:8 ratio in nearly all of the transgenic events containing ChFatB2 TE alone fluctuates between 3 and 6, whereas in the F1 generation of transgenic containing both the TE and the KAS A-2-7, the ratio can be as high as 22. This increase in C:10 levels is accompanied by an increase in the total C:8 and C:10 content (Figure 16). The sum of the C:8 and C:10 fatty acids in the heterozygous F1 lines is as high as those in the homozygous parent line (4804-22-357), whereas the

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heterozygous lines usually contain substantially less C:8 and C:10 than the homozygous lines.

Similar results were observed in F1 generation seeds resulting from crosses performed between 4804-22-357 (ChFatB2) and the 5413-17 event (CpFatB1 and chKAS A-2-7 tandem). Levels of C:8 and C:10 in the 5413-17 line were 6.3 and 2.8 mol% respectively. Data presented in Figure 17 show that there is shift towards C:10 fatty acids as was observed with the 4804-22-357 (ChFatB2) x 5401-9 (chKAS A-2-7) crosses. Furthermore, Figure 18 indicates the presence of two separate populations of heterozygotes. Those containing approximately 9-11 weight percent C:10 + C:8 are believed to represent offspring containing a single copy of the ChFatB1 TE gene and no copies of the CpFatB1 and chKAS A genes from 5413. Those plants containing approximately 15-20 weight percent C:10 + C:8 are believed to represent the heterozygotes containing a single ChFatB1 TE gene as well as the CpFatB1 and chKAS A genes from 5413. Thus, the level of the C:10 + C:8 fatty acids does not decrease to 50% of that detected in parent lines when a copy of the ChKAS A gene is 20 present.

To further characterize the chain length specificity of the Cuphea hookeriana KAS A enzyme, crosses between transgenic Brassica napus lines containing a California Bay (Umbellularia californica) 12:0 specific thioesterase, Uc FatB1 (USPN 5,344,771) and chKAS A-2-7 (5401-9) were made. Half seed analysis of transgenic plants containing Uc fatB1 have previuosly indicated that these plants can accumulate up to 52 mol% C12:0 in the seed oil of homozygous dihaploid

lines (LA86DH186). Crosses between the line LA86DH186 and untransformed control *Brassica* demonstrated a decrease in the C12:0 levels.

However, crosses between LA86DH186 and the 5401-9

hemizygous line led to an accumulation of up to 57 mol%

C12:0 in the seed oil of F1 progeny (Figure 19).

Interestingly, in crosses with LA86DH186 x untransformed control line and LA86DH186 x 5401-9, levels of C14:0 in the seeds of the F1 progeny decreased to 50% of the levels

obtained in homozygous LA86DH186 lines (Figure 20).

Furthermore, increases in the proportion of C12:0 fatty acid resulted in a substantial decline in the proportions of all the long-chain fatty acyl groups (C16:0, C18:0, C18:2, and C18:3). These results indicate that the ChKAS A-2-7 is an enzyme with substrate specificity ranging from C6:0 to C10:0-ACP, and that its over-expression ultimately reduces the longer chain acyl-ACP pools.

Further evidence is obtained in support of the chain length specificity of the ChKAS A-2-7 in crosses of the 5401-9 line with a transgenic line (5266) expressing an 18:1/18:0 TE from Garcinia mangostana (GarmFatA1, US patent application No. 08/440,845). Transgenic Brassica line 5266 has been shown to accumulate up to 24 mol% C18:0 in the seed oil of homozygous lines (Figure 21). However, in the seed oil of F1 progeny of crosses between 5266 and 5401-9 levels of C18:0 were reduced to approximately 12 mol%. Furthermore, levels of C16:0 generated from these crosses was similar to the levels obtained from the seed oil of nontransgenic control plants.

#### Example 5 In vitro Analysis of Plant KAS Enzymes

Seed extracts were prepared from developing seeds of nontransgenic controls or transgenic Brassica expressing chKAS A-2-7 as described in Slabaugh et al. (Plant Journal, 5 1998 in press) and Leonard et al. (Plant Journal, 1998, in press). In vitro fatty acid synthesis assays were performed as described by Post-Beittenmiller (J. Biol. Chem. (1991), 266:1858-1865). Extracts were concentrated by ammonium sulfate precipitation and desalting using P-6 columns (Bio-Rad, Hercules, CA). Reactions (65µ1) contained 0.1M Tris/HCl (pH 8.0), 1 mM dithiothreitol, 25 mM recombinant spinach ACP1, 1 mM NADH, 2 mM NADPH, 50 µM malonyl-CoA, 10  $\mu$ M [1-14C]acetyl-CoA (50 mCi/mmol), 1mg/ml BSA, and 0.25 mg/ml seed protein. Selected seed extracts were preincubated with cerulenin at 23°C for 10 min. Reaction products were separated on an 18% acrlamide gel containing 2.25M urea, electroblotted onto to nitrocellulose and quntitated by phosporimaging using Image QuaNT software (Molecular Dynamics, Sunnyvale, CA). Authentic acyl-ACPs were run in parallel, immunoblotted and finally detected by 20 anti-ACP serum to confirm fatty acid chain lengths.

The results (Figure 22) indicate that the fatty acid synthesis capabilities of transgenic Brasica (5401-9) seed extracts was greater than that obtained from in the nontransgenic controls as measured by the relative abundance of C8:0- and C10:0-ACP at all time points tested. In addition, pretreatment of the extracts with cerulenin, markedly reduced the synthesis of longer chain fatty acids in both the transgenic and nontransgenic control seed

extracts. However, the extension of the spinach-ACP was much less inhibited in the seed extracts from the transgenic lines than in the seed extracts of nontransgenic control Brassica.

These data further support that Ch KAS A-2-7 is a condensing enzyme active on medium chain acyl-ACPs, and that expression of this enzyme in plants results in enlarged substrate pools to be hydrolyzed by medium-chain specific thioesterases. Furthermore, these data suggest that chKAS

A-2-7 also is a cerulenin-resistant condensing enzyme.

All publications and patent applications mentioned in this specification are indicative of the level of skill of those skilled in the art to which this invention pertains.

- 15 All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.
- Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claim.

WO 98/46776

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MISSING UPON TIME OF PUBLICATION

- 13. The construct of Claim 5 wherein said encoding sequence is cpuKAS A/p8-9A.
- 14. The construct of Claim 5 wherein said encoding sequence is chKASIII-27.
- 15. An improved method for producing medium-chain fatty acids in transgenic plant seeds by expression of a plant medium-chain thioesterase protein heterologous to said transgenic plant,

the improvement comprising expression of a plant synthase

10 factor protein heterologous to said transgenic plant in

conjunction with expression of said plant medium-chain

thioesterase, whereby the percentage of medium-chain fatty

acids produced in seeds expressing both a plant synthase factor

protein and a plant medium-chain thioesterase protein is

- increased as compared to the percentage of medium-chain fatty acids produced in seeds expressing only said plant medium-chain thioesterase protein.
- 16. The method of Claim 15 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 20 17. The method of Claim 15 wherein said medium-chain thioesterase protein is a CpFatB1 protein.
  - 18. The method of Claim 15 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.
- 19. The method of Claim 15 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
  - 20. The method of Claim 19 wherein said synthase factor A protein is from a Cuphea species.

- 21. The method of Claim 20 wherein said Cuphea species is C. hookeriana or C. pullcherrima.
- 22. A method of altering the medium-chain fatty acid composition in plant seeds expressing a heterologous plant medium-chain preferring thioesterase, wherein said method comprises

providing for expression of a plant synthase factor
protein heterologous to said transgenic plant in conjunction
with expression of a plant medium-chain thioesterase protein

10 heterologous to said transgenic plant, whereby the composition
of medium-chain fatty acids produced in said seeds is modified
as compared to the composition of medium-chain fatty acids
produced in seeds expressing said plant medium-chain
thioesterase protein in the absence of expression of said plant

15 synthase factor protein.

- 23. The method of Claim 22 wherein said medium-chain thioesterase protein is a ChFatB2 protein.
- 24. The method of Claim 22 wherein said medium-chain thioesterase protein is a CpFatB1 protein.
- 25. The method of Claim 22 wherein said medium-chain thioesterase protein is a C12 preferring thioesterase from California bay.

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- 26. The method of Claim 22 wherein said plant synthase factor protein is expressed from a construct according to Claim 1.
- 27. The method of Claim 26 wherein said synthase factor A protein is from a Cuphea species.
- 28. The method of Claim 27 wherein said Cuphea species is C. hookeriana or C. pullcherrima.

- 29. The method of Claim 22 wherein said fatty acid composition is enriched for C10 fatty acids.
- 30. The method of Claim 22 wherein said fatty acid composition is enriched for C12 fatty acids.
- 31. The method of Claim 22 wherein said fatty acid composition is enriched for at least one medium chain fatty acid and at least one other medium chain fatty acid is decreased.
- 32. The method of Claim 31 wherein said enriched fatty 10 acid is C12 and said decreased fatty acid is C14.

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48	96	144	192	240	288	336	384
GGC Gly	AAG Lys	GGT Gly	CAC His	666 G1y	TCA Ser	GCT Ala	ACT Th <i>r</i>
CCG Pro	TCC	GGT	GGT G1y	ATG Met	$\mathtt{TAT}$	GCC	66C 61Y
CCC	CTC	ATG Met	AAG Lys	AAC	AAC Asn	GCT	GGA Gly
GAT Asp	CGC	GGA Gly	GAG Glu	ACA Thr	CCA	CAT	GCT
GTG Val	GAC	ACA	ATC Ile	ATT Ile	66C Gly	TTC	ATT
CTA Leu	GCC	GGA Gly	CTT Leu	GCC Ala	ATG Met	TGC	ATG Met
GAA Glu	GGT Gly	GTC Val	TCT Ser	TAT Tyr	CTC	$\mathtt{TAC}\\ \mathtt{TY}_{\mathcal{I}}$	CTT
CTA Leu	CTC	CTG	CAG Gln	CCC	GGT Gly	AAC Asn	GAT Asp
GCT Ala	GAT Asp	GTG Val	Grr Val	ATC Ile	TTT Phe	TCC	GCT
GCC	GCC Ala	GGA Gly	666 G1y	TTC Phe	GAA Glu	ACT Thr	GAG Glu
GCG Ala	CGA Arg	GCC	GAC Asp	TTC	ATC Ile	GCC Ala	$_{\rm GGT}^{\rm GGT}$
GTG Val	GCA Ala	AGA Arg	TCT Ser	CCT	GCT	TGT Cys	CGT
GCG Ala	TCG	GAG Glu	TTC Phe	ACC Thr	CTC	GCA Ala	CGC Arg
ACC Thr	AAT Asn	AAG Lys	GTC Val	ATC Ile	CTG	ACT Thr	ATC Ile
TCC	AGG Arg	GAC Asp	ACT	AAA Lys	GCC Ala	TCC	CAT His
AGC Ser	TGC Cys	ATC	CTG	CGG Arg	TCT Ser	ATT Ile	AAT Asn

FIGURE 1 1 OF 4

						•
480	528	576	624	672	720	768
TGG Trp	TTG Leu	ATT Ile	ACT Thr	AGC Ser	GCT Ala	ATC Ile
CCC	GTG Val	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala
AGG Arg	GGA Gly	CCG	CAC His	GAG Glu	ATA Ile	AAT Asn
TCT Ser	GCT Ala	GCA Ala	тат Туг	ATT Ile		ATA
GCC	GGT Gly	GGA Gly	GCT Ala	TGC Cys	AAT Asn	GAG
ACT	GAA Glu	CGA Arg	GAT ASD	TCT Ser	GTC Val	GCC Ala
CAG Gln	GGT G1Y	AGA Arg	TGT Cys	TCT Ser	GAG Glu	CTC Leu RE 1
CCG	ATG Met	ATG Met	AAC Asn	GTC Val	GAA Glu	GAT CTC ASP Leu FIGURE 2 OF 4
GAC Asp		GCA	ATC Ile	GGT Gly	CCT	666 617
GAT Asp	TTT Phe	CAT His	GCA Ala	CTT	TCA	GCT Ala
AAC Asn	GGT Gly	GAA Glu	GGT Gly	$_{\rm G1Y}^{\rm GGT}$	GTC Val	CTA
AGG Arg	GAT Asp	TTG	GGA Gly	GAT Asp	66C Gly	ACT
CAA Gln	CGT Arg	AGC Ser	TTG	GCT Ala	GCT Ala	TCT Ser
TCT	GAC Asp	GAG Glu	TAT Tyr	AGG Arg	gat Asp	ACT Thr
TTG	AAA Lys	ATG Met	GAG Glu	CCA	GAA Glu	GCG Ala
GCT Ala	GAT Asp	GTG Val	GCA Ala	GAT Asp	CTT	CAT His
	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu ATG GAG AGC TTG GAA CAT GCA ATG AGA CGA GGA GCA CCG ATT ATT Met Glu Ser Leu Glu His Ala Met Arg Arg Gly Ala Pro Ile Ile	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu ATG GAG AGC TTG GAA CAT GCA ATG AGA CGA GGA GCA CCG ATT ATT Met Glu Ser Leu Glu His Ala Met Arg Arg Gly Ala Pro Ile Ile GAG TAT TTG GGA GGT GCA ATC AAC TGT GAT GCT TAT CAC ATG ACT Glu Tyr Leu Gly Gly Ala Ile Asn Cys Asp Ala Tyr His Met Thr	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu ATG GAG AGC TTG GAA CAT GCA ATG AGA CGA GGA GCA CCG ATT ATT Met Glu Ser Leu Glu His Ala Met Arg Arg Gly Ala Pro Ile Ile GAG TAT TTG GGA GGT GCA ATC AAC TGT GAT GCT TAT CAC ATG ACT Glu Tyr Leu Gly Gly Ala Ile Asn Cys Asp Ala Tyr His Met Thr CCA AGG GCT GAT GGT CTT GGT CTT TCT TGC ATT GAG AGT AGC Pro Arg Ala Asp Gly Leu Gly Val Ser Ser Cys Ile Glu Ser Ser	TTG TCT CAA AGG AAC GAT GAC CCG CAG ACT GCC TCT AGG CCC TGG Leu Ser Gln Arg Asn Asp Asp Pro Gln Thr Ala Ser Arg Pro Trp AAA GAC CGT GAT GGT TTT GTG ATG GGT GAA GGT GCT GGA GTG TTG Lys Asp Arg Asp Gly Phe Val Met Gly Glu Gly Ala Gly Val Leu ATG GAG AGC TTG GAA CAT GCA ATG AGA CGA GGA GCA CCG ATT ATT Met Glu Ser Leu Glu His Ala Met Arg Arg Gly Ala Pro Ile Ile GAG TAT TTG GGA GGT GCA ATC AAC TGT GAT GCT TAT CAC ATG ACT Glu Tyr Leu Gly Gly Ala Ile Asn Cys Asp Ala Tyr His Met Thr CCA AGG GCT GAT GGT CTT GGT GTC TCT TCT TGC ATT GAG AGT AGC Pro Arg Ala Asp Gly Leu Gly Val Ser Ser Cys Ile Glu Ser Ser Glu Asp Ala Gly Val Ser Pro Glu Glu Val Asn Tyr Ile Asn Ala

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816	864	912	096	1008	1056	1116	
AAG AAG GTT TTC AAG AAC ACA AAG GAT ATC AAA ATT AAT GCA ACT AAG Lys Lys Val Phe Lys Asn Thr Lys Asp Ile Lys Ile Asn Ala Thr Lys	TCA ATG ATC GGA CAC TGT CTT GGA GCA TCT GGA GGT CTT GAA GCT ATA Ser Met Ile Gly His Cys Leu Gly Ala Ser Gly Gly Leu Glu Ala Ile	GCG ACT ATT AAG GGA ATA AAC ACC GGC TGG CTT CAT CCC AGC ATT AAT Ala Thr Ile Lys Gly Ile Asn Thr Gly Trp Leu His Pro Ser Ile Asn	CAA TTC AAT CCT GAG CCA TCG GTG GAG TTC GAC ACT GTT GCC AAC AAG Gln Phe Asn Pro Glu Pro Ser Val Glu Phe Asp Thr Val Ala Asn Lys	AAG CAG CAA CAC GAA GTT AAC GTT GCG ATC TCG AAT TCA TTC GGA TTT Lys Gln Gln His Glu Val Asn Val Ala Ile Ser Asn Ser Phe Gly Phe	GGA GGC CAC AAC TCA GTC GTG GCT TTC TCG GCT TTC AAG CCA TGATTA Gly Gly His Asn Ser Val Val Ala Phe Ser Ala Phe Lys Pro	CCCATTTCAC AAGGTACTTG TCATTGAGAA TACGGATTAT GGACTTGCAG AGTAATTTCC	FIGURE 1 3 OF 4

1470	1296	1348
4794911194	TGAAATTATA TTTATTTTAT CTTAGAAAGG TCAAATAAGA TTTTGTTTTA CCTCTGTAAA	AA
זורופורוו	TTTTGTTTTA	AAAAAAAA
ICCCITINA	TCAAATAAGA	TCTCAAAAAA
ALTALIAMIT	CTTAGAAAGG	GGAAGTGCCG
AAAACTAAGG	TTTATTTAT	GTATTGGAAA
TCTATGIAAT AAAACTAAGG ATTATTAATT ICCCTTTTAA ICCTGTCTCC AGTITGAGCA 1230	TGAAATTATA	ACTITIGITI GIATIGGAAA GGAAGIGCCG ICICAAAAAA AAAAAAAA AA

FIGURE 1 4 OF 4

Sequence Range: 1 to 1704

40 GTG Val>		GCA Ala>		TCT Ser>	0	GAC Asp>	240	cGG Arg>	CTC Leu>		GAA Glu>
GNG		TCG	140	GAC ASP	190	ATC Ile		ATC	AGG Arg		CTC
ACC Thr	90	AAT Asn	7	GTC Val		TTA Leu		CAG Gln	AGG Arg	330	GCT
30 TCC Ser		AGG Arg		GAC Asp		AGC	230	GGC Gly	280 GAC AGG ASP Arg		AAG Lys
AGC		TGC	3.0	TCC	180	ATC Ile	(1	GGC Gly	AAC		AAG Lys
TGG	80	GGC Gly	13	GGC		GGG		TTC	AAG Lys	320	666
20 AGC Ser		CCG		${ t TTC}$		AGC	220	AGG Arg	270 GGG Gly	(F)	GCC
aaa Lys		CCC Pro		GTA Val	170	GAG Glu	22	ACC	GAC Asp		GTC
AAC Asn	70	GAT Asp	120	TCC	•	GGC Gly		CCC	ATC Ile	0,	ATT Ile
10 AAA GGG Lys Gly	•	GTG Val		GTC		TCC		TTC	260 TAC TYF	310	TGC
AAA Lys		CTA		CTC	160	CTC	210	AAG Lys	2 GGA Gly		TAC
ACT		GAA Glu	110	GGC Gly	1(	CTC		TCC	ACG Thr		CGC Arg
CTC	09	CTA	, ,	ATG Met		AAG Lys		GCT	50 GCG Ala	300	CTC
ACC		GCT		GGC Gly		GAA Glu	200	GAC	250 AAC G( Asn A]		TGC Cys
TTA		GCC	100	GCC	150	TAC	•	TTC Phe	TTC		GAT Asp
AAA Lys	20	GCG	Ä	CGA Arg		TAT Tyr		CGC Arg	GGA G1y	90	GAC Asp

FIGURE 2 1/5

FIGURE 2 2/5

	AGA Arg>	0.0	TCT Ser>	480	CCG Pro>	GCC Ala>		TGT Cys>		CGA Arg>	0	ATT Ile>
380	GAG Glu	43	TTC		TCC	CTT Leu		GCA Ala	620	CGC Arg	670	ATC Ile
	AAG. Lys		GTC Val		ATC Ile	20 CTG Leu	570	ACT Thr	v	ATC Ile		GCA Ala
	GAT Asp		ACC Thr	470	AAG Lys	520 GCT CTG Ala Leu		TCA		CAT His		GCT
370	ATT Ile	420	CTA	•	CGG Arg	TCT		ATT Ile	610	AAT Asn	* 099	GAG Glu
m	AAG Lys		66C 61y		CAC His	666 61y	260	TAT TCG Tyr Ser	61	GCC		ACT Thr
	TCC		$_{\rm GLY}^{\rm GGT}$	460	GGT Gly	510 ATG Met	u,	TAT Tyr		GCT Ala		GGA Gly
	AGC CTC Ser Leu	410	ATG Met	4	AAA Lys	AAC Asn		CCA AAC Pro Asn		GCC Ala	650	GCT GGA Ala Gly
360			GGT Gly		GAG Glu	ACA Thr	550	CCA Pro	009	TAT Tyr	Ψ.	
	GAA Glu		ACT		ATC Ile	500 ATT Ile	5,	GGC		TTT Phe		ATT Ile
	GGT Gly	400	GGA Gly	450	CTC	GCC Ala		ATG Met		TGC	0.1	ATG Met
350	GGC Gly	4	GTT Val		AAT Asn	тат Туг		CTG	290	TAC Tyr	640	CTC
	CTC		CTA		CAG Gln	490 ATT CCC Ile Pro	540	GGT Gly	u,	AAC Asn		GAC
	GAT		GTG Val	440	GrT Val			TTG		TCC		GCT
340	TCC	390	GGA Gly	•	$_{\rm GGG}$	TTC		GAT	580	ACT Thr	630	GAG Glu
n	AAT Asn		GCT Ala		GAC	TTT Phe	30	ATC	28	GCT Ala		GGC Gly

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720	AGG Arg>	GAT Asp>		TTG Leu>		GGA Gly>	0	GAT Asp>	096	GGG Gly>	ACT Thr>
	CAA Gln	CGT Arg		AGC	860	TTG	910	GCT		GCT	TCC Ser
	TCT	760 AAG GAC Lys Asp	810	GAG Glu	ω	TAT		AGG Arg		GAT Asp	O ACT Thr
710	TTA	-		ATG Met		GAA Glu		CCA Pro	950	GAA	1000 GCG A( Ala T
•	GCT	GAT Asp		GTT Val	850	GCA Ala	900	GAT Asp	0.	CTG Leu	CAT His
	AGG Arg	TGG Trp	800	TTG Leu	80	ATT Ile		ACT Thr		AGT	GCT
700	TGC	750 CCG Pro	~	GTA Val		ATT Ile		ATG Met	940	AGC	990 AAT Asn
7(	GCC Ala	AGG		GGA Gly		CCG Pro	890	CAT His	6	GAG Glu	ATA Ile
	GTT Val	TCA	790	GCT Ala	840	GCG Ala	w	TAT Tyr		ATT Ile	TAC
	TTC	740 GCC Ala	7.5	$_{\rm GGG}$		$_{\rm GGA}^{\rm GGA}$		GCT Ala		TGC	980 AAT Asn
069	GGA Gly	ACT Thr		GAA Glu		CGA Arg	880	GAT Asp	930	TCT	g GTC Val
	GGA Gly	CAG Gln		GGC	830	AAA Lys	88	TGT		TCC Ser	GAG Glu
	TTA	730 AC CCT SP Pro	780	ATG Met	w	ATG Met		AAT Asn		GTC Val	970 CCT GAA Pro Glu
680	GGG G1y	GA		GTG Val		GCA Ala		GTC	920	$_{\rm GGT}$	97 CCT Pro
Ū	ATT Ile	GAT		TTT Phe	820	CAT His	870	GCA	01	CTT Leu	TCA
	CCA	AAT Asn	10	$_{\rm G1y}^{\rm GGT}$	88	GAA Glu		GGT Gly		$\frac{\text{GGG}}{\text{G1}\text{y}}$	GTC Val

FIGURE 2 3/5

FIGURE 2 4/5

	AAG Lys>		CAC His>	0.0	GGA Gly>	1200	GAG Glu>	GAA Glu>		TCA Ser>		GCA
	TTC Phe	1100	GGA Gly	1150	AAG Lys	77	CCC	CAT		AAC Asn	1340	AAT
1050	GTT Val	11	ATC Ile		ATT Ile		AAT Asn	CAA Gln	1290	CAC His	13	TCA AAT
• •	AAG Lys		ATG Met		ACA Thr	1190	TTC	1240 CAG CAA Gln Gln	1-1	GGC Gly		GGT
	AAG Lys	90	TCG	1140	GCG Ala	7	CAA Gln	AAG Lys		GGA Gly	0	
1040	ATC Ile	1090	AAG Lys	•	ATT Ile		AAC Asn	AAG Lys	1280	TTC	1330	TTA CTC
Ä	GCC		ACT Thr		GCC Ala	30	ATA Ile	1230 F GCC AAC P I Ala Asn I	13	GGA Gly		TGA
	AAT Asn		AAT GCA Asn Ala	1130	GAA Glu	1180	AGC	GCC Ala		TTC Phe		CCA
30	ATA Ile	1080	AAT Asn	Ħ	CTT		CCC	GT	0.2	TCA	1320	TTC AAG Phe Lys
1030	GAG	•	ATC Ile		GGT		CAT His	1220 GAC ACA Asp Thr	1270	AAT Asn	-	TTC
	GCC		ACA Thr	20	$_{\rm GGG}$	1170	CTT	12 GAC ASP		TCA		GCC
	CTT Leu	1070	ATC Ile	1120	TCA	,,	TGG Trd	TTC Phe		ATC Ile	1310	TCA
1020	GAT Asp	Ä	GAA Glu		GCA		GGC Gly	10 GAA G1u	1260	GCT Ala	ਜ	TTC
	666 G1y		AAG Lys		GGA Gly	1160	ACC Thr	1210 A GTG GA r Val Gl	• •	GTT Val		GCT
	GCT Ala	09	ACC	1110	CTT Leu	H	AC( Th:	J.C.		AAT Asn	00	GTA Val
10	CTT Leu	1060	AAC Asn	• •	TGT		ATA Ile	CCA '	20	GTG Val	1300	GTT

SUBSTITUTE SHEET (RULE 26)

FIGURE 2 5/5

AATTIGITGC IGAGACAGIG AGCITCAACT IGCAGAGCAA ITITITACAI GCCITGICGI CGGAAGAGCG TAATACCGGG ATAGTTCCTT GATAGTTCAT TTAGGATGTT TTACTGCAAT AATCGAAGAT TATTTCCATT CTAATCCAGT CTCCGNCGAG TTTGAGAATC TATCTGTTTG TATTAGAAAG AACGAGGCAA GATTTTGTTT CATGTTTGTG TTTGTATTAC TTTCTTTTTG CCCTTGTCAA TGGCATTTAA GATAAGCTTA TAAAAAAAA AAAAAAAAA AAAACTCGAG GGGGGGCCCG GTACCCAATT CGCCCTATAG TGAGTCGTAT GACAATTCAC TGTCCGTCGG

1/6	
3	
JRE	
FIGI	

09*	CCGCTCTAGA ACTAGTGGAT	120	GGTCGGCTCA GCTCAGGTGT	ACG TGG Thr Trp		CGT TCC Arg Ser	0	CTC TCC Leu Ser	310	CCT TGC Pro Cys	360	TTC GGA Phe Gly
20	GA AC	110	CA GC	TGT Cys	210	GAC CCA CGT Asp Pro Arg	260	ACT OTHE	. (*)	GAT C Asp I		CTC
	TCTA	Н	GGCT	160 TTC	7	GAC		AGG		CTC	0	GCT TCC (Ala Ser 1
				C CCT		AAC Asn		CGG Arg	300	CAA TGC Gln Cys	350	GCT
40	GAGCTCCACC GCGGTGGCGG	100	TTCTTACTTG	G TCC (	200	GAC	250	CGC	•••			TTC Phe
	GGTG		CTTA	140 TCT TGC ATG GTT GCG Ser Cys Met Val Ala	7	TCC		CGT		TTC		GGA Gly
0	ည	0		1 3 GTT E Val		TCA		TCC	290	ACC Thr	340	AAC Asn
30	CCAC	90	SAGT	c ATG s Met		ACT	240	CGC CTC Arg Leu	73	TCC		GAT Asp
	AGCT		GGCACGAGTT	140 T TGC r Cys	190	CCC	••			CTC CGC GGA Leu Arg Gly		TTC CTC GGG Phe Leu Gly
20	rg G	80		14 r TCT a Ser		ATG Met		CTC		CGC Arg	330	CTC
•	AAGC	~	SAAT	c GCT r Ala		TGC ATG Cys Met 1	230	CGG Arg	280		(-1	TTC
	ACTAAAGGGA ACAAAAGCTG		CCCCCGGGCT GCAGGAATTC	o G ACC a Thr	180	GCT GCA Ala Ala	.2	AAG Lys		TCC		CGC Arg
10	GGA	70	GCT	130 3 GCG t Ala				CAC		TGC	320	CAA
	AAAG		CCGG	A ATG Met		GTA Val		TCC	270	CAT His	ä	CAG Gln
	ACT		CCC	TCCA	170	CTC	220	CTT Leu	- •	TCC		AAC Asn

ACT Thr		GAA Glu		GTG Val		TAC	009	AAC	TCT		GAC
CGC	•	CAG Gln	200	GTT Val	550	GTT Val	Ψ	GAG Glu	AAG Lys		ATG
66C 61y	450	GCA Ala	2(	GTA Val		GAT Asp		ATA Ile	ATC	069	AGG Arg
400 CTC Leu	•	CCT		CGA Arg		CCC	290	GAG Glu	640 GAG Glu	v	GAG Glu
AGG Arg		CAA Gln		AGG	540	GAC	55	AGT	GGA Gly		TCC
CTG	440	ATG	490	CAA Gln	۵,	CAT His		ATA Ile	GCC	680	TTC
390 CAC His	44	GCT		AAG Lys		GGC Gly		GGC	630 ATT Ile	99	AAG Lys
66C G1y		GTG Val		ACC	30	CTA	580	AGT	AGA Arg		CCA
CGC		GCT	480	GCT	53	CCT	٠	ATA Ile	ACG		GCC
380 TCA AAȚ Ser Asn	430	ATG	•	CCT		ACT		GAC GGA ASP GIY	620 T CCC	049	GTG Val
		GTC Val		aaa Lys		GTG Val	570	GAC	62 TTT Phe		TGG Trp
CGT		GAG Glu	470	AAG Lys	520	GTG Val		CTA	CAG Gln		GGC Gly
CTT Leu	420	GGG Gly	4.7	AAT Asn	`	GGC Gly		CTC	TCT Ser	¢60 *	GAT
370 CCT Pro	•	TCC		ACA Thr		ATG Met	260	AAT Asn	610 TGC Cys	v	ACA Thr
AAG Lys		CAT His		TCC	510	GGT	26	AAC Asn	GAC Asp		TCC
TCC	410	TCC	460	GTC Val	υ,	ACA Thr		TAC	TTC	650	TTT Phe

FIGURE 3 2 OF 6

0	GAT Asp		TGT	840	GAT Asp	TGT Cys		GAC		ACA Thr		GAA Glu
740	GCA	790	AAG Lys	w	AGC	TTT Phe		ATG Met	980	GCA Ala	1030	GGC Gly
	TTA		AGA Arg		TTC	CCC	930	GCA Ala	96	TGT		AAA Lys
	GCA Ala		aaa Lys	30	GTA Val	880 AGT Ser	01	CTT Leu		GCC Ala		ATC Ile
730	AAA Lys	780	AAT	83	AAG Lys	ATC Ile		ATT Ile		ACT Thr	1020	ATA Ile
·	AAG Lys	•	CTC		ATG Met	AAG Lys	920	GCT	970	TCA	7(	CAC His
	GGC Gly		GAG Glu		GGT Gly	870 AAG Lys	6	TCC		ATA Ile		AAC
720	GCA Ala	770	AAA Lys	820	GGC Gly	TAT Tyr		GGA Gly		TCG	10	GCG Ala
7	ACT	7.	ATG		TTG	TCA		ATG Met	096	₽₽	1010	GCT
	CTG		GCG		GGA Gly	860 AGG ACT Arg Thr	910	AAT	01	AAC Asn		AAT
0	* ATG Met		GAT	810	TCC	8 AGG Arg		ACA Thr		CCT		CTG
710	TAC Tyr	760	GAA Glu		GGC Gly	CTG		ACC	950	GGC Gly	1000	ATA Ile
	CTT		ACT		ATT Ile	GCT Ala	006	TCT	9,	ATG Met	• • •	TGT Cys
	ATG		ATC Ile	800	CTC	850 GAA Glu	O,	TTT Phe		TGG Trp		TTC
700	TTC	750	GGA Gly	æ	GTT Val	ATT Ile		CCT Pro		GGA Gly	066	AAC Asn
٠	AAG Lys	•	GGT Gly		GGA Gly	TCC	890	GTA Val	940	TTG	O1	AGT

FIGURE 3 3 OF 6

1080	GTT Val	AAT Asn		TTT Phe		CAT His		AGT	1320	O K	TCG Ser
1(	CCT	AAT Asn		GAT GGA Asp Gly	0.	GAG Glu	1270	GGG	13	GGA Gly	GGA GTC Gly Val
	TTA Leu	agg Arg	1170.		1220	TTA	М	GGT		GAA Glu	GGA Gly
20	GTT Val	1120 CAG	H	CGT		GAG Glu		CTA	01	CCT Pro	1360 CAG TCC Gln Ser
1070	GCC	TCA		AAT Asn		GAG	1260	TTT Phe	1310	CAC	cAG Gln
	GCG Ala	TTG	20	GAC AGT Asp Ser	1210	CTT	7	GAA Glu		CCT	GCT
	GAT	1110 CGA GCT Arg Ala	1160	GAC Asp	••	CTT		GCG Ala		GAG Glu	1350 GCC TTG Ala Leu
1060	TCG			TGG		TTA	20	ATT TAT Ile Tyr	1300	ATG ACC Met Thr	GCC Ala
	6GC G1y	TGC		AGA CCA Arg Pro	1200	GTT Val	1250	ATT	` '	ATG	AAG Lys
	GGT Gly	1100 GTA GCA Val Ala	1150	AGA	ä	GGA Gly		ACC		CAC His	1340 ATA GAG Ile Glu
1050	TGT			TCG		GCT Ala		GCA Ala	1290	TAC	
7	CTT	TTC		GCT	06	GGA Gly	1240	GGT Gly	ï	GCC	TGC
	ATG	GGT Gly	1140	aaa Lys	1190	GAA Glu	••	AGA Arg		GAC	CTC Leu
1040	* ATG Met	1090 GGA Gly	H	ACC		GGA Gly		AAA Lys	30	ACT TGC Thr Cys	1330 ATC Ile
10	GAC	TTG		CCT		ATG Met	1230	AAG Lys	1280		1 GTG Val
	GCA Ala	GGT Gly	1130	GAC	1180	GTG Val	17	GCA Ala		TTC	GGT

FIGURE 3 4 OF 6

1410

1400

1390

1380

1370

GCT		AAC		CTT	1560	AGG	66C 61y		GTC		TCC
ACT TCC ACT CCT GCT Thr Ser Thr Pro Ala	20	CAA Gln	1510	CAC CTT CTT His Leu Leu	1.5	ATA Ile	GAA Glu		AAG Lys	00	TCA
ACT	1460	GGC Gly	•			GCA Ala	1600 GAC CCG GAC ASP Pro ASP	1650	CTG	1700	AAC Asn
TCC		TTC		$_{\rm G1y}$	0.0	CAG Gln	1600 CCG	ï	AAA Lys		CAT His
ACT Thr		TGT	1500	ATC Ile	1550	GTT Val	1 GAC ASP		GAG Glu		GGC Gly
CAT GCA His Ala	1450	CAC	7	ATG Met		GTA Val	GAA Glu	40	AAG Lys	1690	GGC
CAT	•	GCC		TCG		GCA Ala	1590 AAT TTG Asn Leu	1640	AAG Lys	• •	TTC Phe
GCG		CTC	06	AAA Lys	1540	GTT Val	15 AAT Asn		CCT		666 61y
ATA AAT Ile Asn	1440	GCT	1490	ACC	•	GCA Ala	ATT Ile		GGC Gly	1680	TTT Phe
ATA Ile	À	CAA Gln		TCC		GAA Glu	1580 CCA AAT Pro Asn	1630	GTC	Ä	AAT TCA Asn Ser
TAC		TAC		AAT Asn	1530	GTA Val		.,	CTC		
AAT	30	GAA Glu	1480	GTG	7	GGT GGC (Gly Gly )	ATC.CAT Ile His		CTG	0.0	TTG TCC Leu Ser
GAC GTA Asp Val	1430	AAG Lys		aga Arg				1620	AAA Lys	1670	
GAC		ATC Ile		CTG	50	GCT	1570 GGA TGG Gly Trp	ī	GCA Ala		GGT Gly
GAA Glu		GAT Asp	1470	GAG Glu	1520	GGA G1y			GAT Asp		GTC Val
AGG Arg	1420	GGA Gly	तं	AGT Ser		GGA Gly	ACA Thr	1610	GTG Val	1660	AAG Lys

FIGURE 3 5 OF 6

1710	1720	1730	1740	1750	1760	
ATA CTA TTT Ile Leu Phe	GCC	CCC TGC AAC TAG A AAAGAGTCTG TGGAAGCCGA GAGTCTTTGA Pro Cys Asn ***	. AAAGAGTCTG	TGGAAGCCGA	. GAGTCTTTGA	
1770	1780	1790	1800	1810	1820	
GAACTCATGC	GAACTCATGC ACGTTAGTAG		CTTCTTATGC CTCTGAAACC GAGATAGACC GGCTACTCGA	SAGATAGACC	GGCTACTCGA	
1830	1840	1850	1860	1870	1880	
GGGGATGCCA	AAGATACTCC	GGGGATGCCA AAGATACTCC TTGCCGGTAT TGGTGTTAAG AGATCACTGC TTGTCCCTTT	TGGTGTTAAG	AGATCACTGC	TTGTCCCTTT	
1890	1900	1910	1920	1930	1940	
TATTTTCTTC	TTCTTTTGAG	TATTITCTIC TICTITIGAG AGCTITAACC GAGGTAGTCG TATTITCGAG	GAGGTAGTCG		CTTTTCGAAT	
1950	1960	1970	1980	1990	2000	
ACATGTTCGT	TATCGGATCA	TATCGGATCA ATGTGTTTCT TCTAAGATCA	TCTAAGATCA	TTTGTAATGC ATATTTTGAA	ATATTTGAA	
2010	2020	2030	2040			
AAACCACATC	TCAGTATGCA	AAACCACATC TCAGTATGCA AAATAAAAA AAAAAAAAAA	AAAAAAAAA	AAAAA		

Sequence Range: 1 to 1921

09	TCACCTCTTA CCTCGCCTGC TTCGAGCCCT GCCATGACTA CTACACCTCC	120	GCATCCTTGT TCGGATCCAG GCCCATCCGC ACCACCCGCA GGCACCGGAG GCTCAATCGA	180	GCTTCCCCTT CCGGGGAGGC AATGGCTGTG GCTCTGCAAC CTGCACAGGA AGTTACCACA	ATG Met>		AAT Asn>		TGT Cys>	370	ACA Thr>
	CTAC!		3CTC2		\GTT2	GGA G1y		AAT Asn	320	GAT TGT ASP Cys	ω Γ	TTC TCC ACA Phe Ser Thr
20	CTA (	110	3AG	170	3GA i	220 GTG ACT Val Thr	270	TAC	,	TTT Phe		
	ATGA(	.,	ACCG		CACA(			TTC		ACC Thr		TCT
	225		GGC,		CTG	GTT Val		GTT Val	310	GAG Glu	360	AAG Lys
40	CCCT	100	CGCA	160	CAAC	GTA Val	260	GAT Asp	m'	ATA Ile		ATC
	CGAG		CACC		rc <sub>TG</sub>	210 CGA Arg	••	CCT		GAG Glu		GAG Glu
0	) TT	0	A AC	0	C C	CGG Arg		GAC		AGT	350	GGA Gly
30	CTG	90	rccg	150	TGT	CAG Gln	250	CAT His	300	ATA AGT Ile Ser	(*)	GCT Ala
	CTCG		CCCA		\TGG(	200 ATC AAA CAG Ile Lys Gln	22	66c 61y		AGT GGC Ser Gly		ACG AGA ATT GCT GGA Thr Arg Ile Ala Gly
20	ra co	80	AG G(	140	3C A	ATC Ile		CTA Leu		AGT	340	aga Arg
••	CTCT	~	ATCC/	7,	SGAGO	AGT		CCT	290	ACG Thr	3,	ACG Thr
	rcacc		rcgg,		3000	190 AAG CCA Lys Pro	240	ACT Thr	(1			CCT
10	₽GG 1	70	rgr :	130	T.L.			GTG Val		GAT Asp		TTT Phe
	CGGCACGAGG		rccr	• •	וכככנ	AAG Lys		GTG Val	280	CTT	330	CAA Gln
	)992		GCA.		GCT	AAG Lys	230	GGT Gly	28	CTG	-	GCT

FIGURE 4

420	ATG Met>	ATC Ile>		CTC Leu>		GAA Glu>	610	TTC Phe>	099	TGG Trp>	TTT Phe>
	TTC	GGA Gly		GTT Val	260	ATT Ile	61	CCT		GGA G1y	AAC Asn
	GAC AAG ASP Lys	50 GGT Gly	510	GGA Gly	.,	GCC		GTA Val		TTG	700 ACG AGT Thr Ser
410	GAC	460 AAT GG1 Asn Gl3		TGC		GAT Asp		$ extstyle{TGT}$	650	GAC	
•	ATG Met	ACA		AAA Lys	550	AAT Asn	¢009	TTT Phe	ŭ	ATG Met	GCA
	AGG	TTA	200	AGA Arg	50	TTC		CCC		GCA	TGT Cys
400	AAG Lys	450 GCA Ala	<b>.</b> ,	AAA Lys		GTA		AAT Asn	640	CTT	690 GCT Ala
4	TCC	AAA Lys		GAT Asp		AAG Lys	290	ATG Met	9	ATG Met	ACT Thr
	CIC	AAG Lys	490	GAG CTA Glu.Leu	540	ATG Met		AAG Lys		GCT Ala	TCT Ser
	CCG AAG Pro Lys	440 GGC Gly	4			GGA Gly		AAG Lys		TCA	680 ATA Ile
390	CCG	GCC Ala		AAA Lys		GGT Gly	30	TAT Tyr	630	GGA	TCG Ser
	GCC	ACT		ATG Met	530	ATG Met	28	TCA		ATG Met	TAC
	GTG Val	30 CTG Leu	480	GTG Val		GCA		ATT Ile		AAT Asn	670 CCC AAC Pro Asn
380	TGG	430 ATG CTC Met Leu		GAT Asp		TCA		AGG Arg	620	ACA Thr	67 CCC Pro
••	GGT	TAC Tyr		GAA Glu	0 2	GGC Gly	570	CTA	Ψ	ACC Thr	GGC Gly
	GAT Asp	CTT	470	ACC Thr	52	ATT Ile		GCC Ala		GCT	ATG Met

FIGURE 4 2/6

	GTG Val>		GGA Gly>	850	ACT Thr>	006	$\frac{\text{GGG}}{\text{G1}\text{y}}>$	AAA Lys>		TGC Cys>		ATT Ile>
	GAT Asp	800	ATG Met	8	CCT		ATG Met	AAG Lys		ACT	1040	GTG Val
750	GCA Ala	w	GGT		GAC		GTT Val	940 T GCA s Ala	990	TTC	7(	GGA Gly
	GAA Glu		ATT Ile		GCC Ala	890	TTT Phe	940 CAT GCA His Ala		AGT		GCT Ala
	GGC Gly	790	CCT Pro	840	AAT Asn	w	GGA Gly	GAG Glu		GGA Gly	30	GGA Gly
740	AGA Arg	75	ATA Ile		AGA Arg		GAT ASP	TTA Leu	980	GGT Gly	1030	GAT Asp
1-	ATC Ile		ATC Ile		CAG Gln	880	CGT Arg	930 GAG Glu	O1	CTA		CCT
	ATA Ile		GTA Val	830	TCA	86	AAT Asn	GAG Glu		$\mathbf{r}\mathbf{r}\mathbf{r}$		CAC His
730	CAC	780	GCG Ala	ω	TTG		AGT	CTA	970	GAA Glu	1020	GAG CCT Glu Pro
73	AAC Asn		GAT Asp		GCT		GAC	920 CTA Leu	9.1	GCA Ala	17	GAG Glu
	GCG Ala		TCA	820	CGA Arg	870	TGG Trp	9 CTA Leu		TAC		ACC Thr
	GCT	170	GGC Gly	8	TGC		CCA	GTG Val		ATT Ile	1010	ATG
720	AAT Asn		GGG		GCA		AGA Arg	910 GCT GGA Ala Gly	960	ACT Thr	1(	CAC
	CTG		TGC		GTT Val	860	TCA	91 GCT Ala		GCG		$\mathtt{TAC}$
	ATC Ile	760	CTT	810	$ ext{TTT}$	w	GCT	GGA Gly		GGT Gly	00	GCC Ala
710	TGT Cys	7(	ATG		GGT Gly		AAA Lys	GAA Glu	950	AGA	1000	GAT ASP

FIGURE 4

1090	GAA GAC Glu Asp>	1140	AT ATC	G TTA u Leu>		GCA GCC Ala Ala>		G IGG Y Trp>	1330	T ACC P Thr>	1380	99
Н			GAT Asp	GAG Glu			1280	. GGG	7	GAT		GTC
	AGG Arg		GGA Gly	1180 AAC AAC Asn Asn	1230	GGA Gly	$\vdash$	ACT		GTG Val		AAG Lys
	TCT	1130	GCT Ala			CTC		AGG Arg		GGC G1y	1370	ATT Ile
1080	GTC Val	H	CCA Pro	CAA Gln		CTT	0	ATA Ile	1320	GAA Glu	13	AAC ATT Asn Ile
-1	GGA Gly		ACT	GGC Gly	1220	CAC His	1270	GCA ATA Ala Ile	Н	GAT Asp		CTG Leu
	TCA	00	TCC	1170 Trc Phe	12	GGT Gly		CAG Gln		CCA	0.0	GAG AGA Glu Arg
1070	CAG	1120	ACA Thr	TGT Cys		ATT Ile		GTT Val	1310	AAC Asn	1360	GAG Glu
1(	GCT		GCC Ala	CAC His	0,	ATG	1260	TCA GTA GTT Ser Val Val	13	GAA Glu		AAG Lys
	TTG		CAT His	160 ATC Ile	1210	TCA	177			TTG		AAG Lys
20	GCT	1110	GCA Ala	1. CTT Leu		aaa Lys		GTT Val	00	AAT Asn	1350	CCT
1060	AAG Lys		AAT Asn	GCT		ACC Thr	1250	GCA Ala	1300	ATT Ile	•	GGC G1y
	GAG		ATA Ile	50 CAA Gln	1200	TCT	17	GAA Glu		AAT Asn		GTG Val
	ATA Ile	1100	TAC Tyr	1150 TAC C TYF G		AAT Asn		GTG Val		CCG	1340	CTC
1050	TGC	H	AAT Asn	GAG Glu		GTG Val	0.4	GGT Gly	1290	CAT His	H	TTG
• •	CTC		GTA Val	AAA Lys	1190	AAA Lys	1240	GGT Gly	1	ATC		AAA Lys

160RE 4

CTC TTC Leu Phe>	1480	TCAAA	1540	CATGCCCATG	1600	GGCGACACAG	1660	TTTCTGAAAT	1720	GAAGAGAACA	1780	TTTATCGCCG	1840	ATCATTGGAG
1420 TCG TCC ATA Ser Ser Ile	1470	CATGTGGA ATTCTACTCA ATCTATCAAA	1530	GCTGAAGTTT TGAGGACTCC AGCATGTTGG TAGCTCCTTA CGTCTCTAGA CATGCCCATG	1590	GAGTACTCAT	1650	TCCCATTTTT	1710	AGTCAGTGAA GAAGAGAACA	1770	TGCTCTCTAT	1830	TTTTGTGGGT TAAAATTTGT AAAACTAGAC GACTGGTTTG TTTTCTCTTG ATCATTGGAG
1410 1420 GGG CAC AAC TCG TCC Gly His Asn Ser Ser	1460	rgrgga arrcı	1520	TAGCTCCTTA	1580	AGTCGGAACC ATGACGGATT	1640	TGTTAGAGCA CTATTCATTA	1700	CGTTTCATCG	1760	CCCTTTGTTT	1820	GACTGGTTTG
1400 GGG TTT GGT ( Gly Phe Gly (	1450	TAG GGCGTTT CATG: ***>	1510	AGCATGTTGG	1570		1630		1690	TACTTTCGAG	1750	TAACCATTTG	1810	AAAACTAGAC
TCA TTC Ser Phe	1440	AAC Asn	1500	TGAGGACTCC	1560	CGGGAGCTGT	1620	TTGCTAGAAT	1680	CTCCCTCCTT ACGGTAGTTG	1740	GGGCACGTAG	1800	TAAAATTTGT
1390 TTG TCT AAT Leu Ser Asn	1430	GCC CCT TAC Ala Pro Tyr	1490	GCTGAAGTTT	1550	AGTTTTGTGT	1610	GATATACTCC	1670	CTCCCTCCTT	1730	AAGCTAACTC	1790	TTTTGTGGGT

FIGURE 4 5/6

			<b>K</b>	A ARREGERE PEREFEREN	dagagaga
				*	
				1920	1910
AAAAAAAAA	AAAAAAAAA	ATAAAAAAA	TTCATTGATG	ATGTATGGCC ATATTTGCCT TTCATTGATG ATAAAAAA AAAAAAAAA AAAAAAAA	ATGTATGGCC
1900	1890	1880	0/8T	1807 1807	000

FIGURE

09	120	169	217	265	313	361	409	457	505
CTGGTACGCC TGCAGGTACC GGTCCGGAAT TCCCGGGTCG ACCCACGCGT CCGTCTTCCC	ACTCCGATCG TTCTTCTTCC ACCGCATCTC TTCTCTTCTC	CGCCGCC ATG CAT TCC CTC CAG TCA CCC TCC CTT CGG GCC TCC CCG CTC Met His Ser Leu Gln Ser Pro Ser Leu Arg Ala Ser Pro Leu 1	GAC CCC TTC CGC CCC AAA TCA TCC ACC GTC CGC CCC CTC CAC CGA GCA Asp Pro Phe Arg Pro Lys Ser Ser Thr Val Arg Pro Leu His Arg Ala 15	TCA ATT CCC AAC GTC CGG GCC GCT TCC CCC ACC GTC TCC GCT CCC AAG Ser Ile Pro Asn Val Arg Ala Ala Ser Pro Thr Val Ser Ala Pro Lys 35	CGC GAG ACC GAC CCC AAG AAG CGC GTC GTG ATC ACC GGA ATG GGC CTT Arg Glu Thr Asp Pro Lys Lys Arg Val Val Ile Thr Gly Met Gly Leu 55	GTC TCC GTT TTC GGC TCC GAC GTC GAT GCG TAC TAC GAC AAG CTC CTG Val Ser Val Phe Gly Ser Asp Val Asp Ala Tyr Tyr Asp Lys Leu Leu 65	TCA GGC GAG AGC GGG ATC GGC CCA ATC GAC CGC TTC GAC GCC TCC AAG Ser Gly Glu Ser Gly Ile Gly Pro Ile Asp Arg Phe Asp Ala Ser Lys 80	TTC CCC ACC AGG TTC GGC GGC CAG ATT CGT GGC TTC AAC TCC ATG GGA Phe Pro Thr Arg Phe Gly Gly Gln Ile Arg Gly Phe Asn Ser Met Gly 95	TAC ATT GAC GGC AAA AAC GAC AGG CGG CTT GAT GAT TGC CTT CGC TAC Tyr lle Asp Gly Lys Asn Asp Arg Arg Leu Asp Asp Cys Leu Arg Tyr 120

553	601	649	697	745	793	841	889
GCC	666 Gly	CTT Leu	GCC Ala 190	ATG Met	TGC Cys	ATG Met	66C G1Y
GGT Gly	GTT Val	TCT Ser	TAT Tyr	CTG Leu 205	TAC	CTT Leu	GGA Gly
CTC Leu 140	CTG Leu	CAA Gln	CCC	GGT Gly	AAC Asn 220	gat Asp	TTG
GAT Asp	GTG Val 155	GTT Val	ATC Ile	CTC	TCC	GCT Ala 235	666 G1y
GCC Ala	GGA Gly	GGG Gly 170	TTC Phe	GAA Glu	ACT	GAG Glu	ATT Ile 250
gac Asp	GCC Ala	GAC ASD	TTC Phe 185	ATT Ile	GCC	GGT Gly	CCA
GAG Glu	AGA Arg	TCT Ser	CCT	GCT Ala 200	TGT Cys	CGT	ATT Ile
CTT Leu 135	GAG Glu	TTC Phe	ACC Thr	CTC	GCA Ala 215	CGC	ATC Ile
TCT	AAG Lys 150	GTC Val	ATC Ile	CTG	ACT Thr	ATC Ile 230	GCA Ala
AAG Lys	gac Asp	ACT Thr 165	AAA Lys	GCC Ala	TCC	CAT His	GCC Ala 245
AAG Lys	ATC Ile	CTG	CGG Arg 180	TCT Ser	ATT Ile	AAT Asn	GAG Glu
666 G1y	AAG Lys	GGT Gly	CAC His	GGG Gly 195	TCA	GCT Ala	ACT
GCC Ala 130	TCC	GGT Gly	GGT Gly	ATG Met	TAT Tyr 210	GCT Ala	GGC
GTC Val	CTC Leu 145	ATG Met	AAG Lys	AAC Asn	AAC Asn	GCT Ala 225	GGA Gly
ATT Ile	CGC Arg	GGA G1y 160	GAG Glu	ACA Thr	CCA Pro	CAT His	GCT Ala 240
TGC Cys	GAC ASP	ACA Thr	ATC Ile 175	ATT Ile	66C Gly	TTC Phe	ATT Ile

FIGURE 5

937	985	1033	1081	1129	1176	1224	1272
ACT Thr 270	GAA Glu	CGA Arg	GAT Asp	TCT Ser	GTC Val 350	GCC Ala	AAA Lys
CAG Gln	GGT Gly 285	AAA Lys	TGT Cys	TCC	GAG Glu	CTC Leu 365	ATC Ile
CCT	ATG Met	ATG Met 300	AAC Asn	GTC Val	GAA Glu	GAT Asp	GAT ASP 380
GAC ASP	GTG Val	GCA Ala	ATC Ile 315	GGT G1Y	CCT	$^{\rm GGG}_{\rm G1Y}$	AAG Lys
GAT Asp	TTT Phe	CAT His	GCA Ala	CTC Leu 330	TCA	GCT Ala	ACA
AAC Asn 265	$_{\rm GGT}^{\rm GGT}$	GAA Glu	GGT Gly	GGT Gly	GTC Val 345	CTA	AAC Asn
agg Arg	GAT ASP 280	TTG Leu	GGA Gly	GAT Asp	GGC Gly	ACT Thr 360	AAG Lys
CAA Gln	CGT Arg	AGC Ser 295	TTG	GCT Ala	GCT Ala	TCT Ser	TTC Phe 375
TCT	GAC	GAG Glu	TAT Tyr 310	agg Arg	GAT Asp	ACT	GTT Val
CTG	aaa Lys	CTG	GAG Glu	CCA Pro 325	GAA Glu	GCG Ala	AAG Lys
GCT Ala 260	GAT Asp	GTG Val	GCA Ala	GAC	CTT Leu 340	CAT His	AAG Lys
AGG Arg	TGG Trp 275	TTG	ATT Ile	ACT Thr	AGC Ser	GCT Ala 355	ATC Ile
TGC Cys	CCC	GTG Val 290	ATT Ile	ATG Met	AGT Ser	AAT Asn	GCC Ala 370
GCT Ala	AGG Arg	GGA Gly	CCT Pro 305	CAC His	GAG Glu	ATA Ile	AAT Asn
GTG Val	TCT	GCT	GCA Ala	TAT Tyr 320	ATT Ile	TAC	ATA Ile
TTT Phe 255	GCC Ala	GGT Gly	GGA Gly	GCT Ala	TGC Cys 335	AAT Asn	GAG Glu

FIGURE 5

1320	1368	1416	1464	1512	1569	1629	1689	1712
ATT AAT GCA ACT AAG TCA ATG ATC GGA CAC TGT CTT GGA GCC TCT GGA Ile Asn Ala Thr Lys Ser Met Ile Gly His Cys. Leu Gly Ala Ser Gly 385	GGT CTT GAA GCT ATA GCG ACT ATT AAG GGA ATA AAC ACC GGC TGG CTT Gly Leu Glu Ala Ile Ala Thr Ile Lys Gly Ile Asn Thr Gly Trp Leu 400	CAT CCC AGC ATT AAT CAA TTC AAT CCT GAG CCA TCC GTG GAG TTC GAC His Pro Ser Ile Asn Gln Phe Asn Pro Glu Pro Ser Val Glu Phe Asp 415	ACT GTT GCC AAC AAG CAG CAA CAC GAA GTT AAT GTT GCG ATC TCG Thr Val Ala Asn Lys Lys Gln Gln His Glu Val Asn Val Ala Ile Ser 435	AAT TCA TTT GGA TTC GGA GGC CAC AAC TCA GTC GTG GCT TTC TCG GCT Asn Ser Phe Gly Phe Gly Gly His Asn Ser Val Val Ala Phe Ser Ala 455	TTC AAG CCA TGA TTACC CATTTCACAA GGCACTTGTC ATTGAGAGTA CGGTTGTTCG Phe Lys Pro 465	TCAAACCCAT TTAGGATACT GTTCTATGTA AAAAAAGTA AGGATTATCA CTTTCCCTTC	TAATCCTGTC TCCAGTTTGA GAATGAAATT ATATTTATTT TAAAAAAAA AAAAAAGGGC	GGCCGCTCTA GAGGATCCAA GCT

FIGURE 5

Sequence Range: 1 to 1802

09	TTATCTCCGC	CCT TCC Pro Ser		TCC Ser	210	CGT Arg		CGG Arg		GTC Val	CTA
	rtatc	CAC TCC CCT His Ser Pro	160	TCC	N	ATC Ile		AAG Lys		GAC	350 ATC AGC Ile Ser
20		CAC 1 His S		CCC		GTC Val		AAG Lys	300	GGC TCC Gly Ser	35 ATC Ile
	CGCGTCCGGG CTTTCCGACC ACATTTCATT TCTTGCCTCG	CTC (		CTC AAT TCC Leu Asn Ser	200	CCC	250	CCC	(-,		GGC
	TCT	100 TCC ( Ser 1	150	AAT Asn	2(	CTC		GAC		${ m TTC}$	AGC
40	CATT	CAA Gln	, ,	CTC		AGC		TCC	290	GTC	340 GAG Glu
	ATTT(			CGC		GCC	240	GAG	55	TCC	66C 61y
0	Z AC	C C	140	TTC	190	CGC Arg	•	CGC		GTC Val	TCC
30	CGAC	80 90 CCGTCGTTCG CCGCCGCCG C ATG Met	17	CCC		CGT Arg		AAG Lys		CTC	330 CTC Leu
	rttc(	2000		GAG Glu		CTC	230	CCC Pro	280	GGC	CTG Leu
20	3G C.	80 CG		CTC	180	CCC	.2	GCC		ATG Met	AAG Lys
•	rccG	CGTT	130	CCT (	•••	CGC		TCC		GGC Gly	320 TAC GAC Tyr Asp
	CGCG'	CCGT		TCC		CTC		GCC	270	ACC	
10	CCA			CCC	170	GCT	220	ACC	••	ATC	TAC
	GGTCGACCCA	70 CGCTCCTCCG	120	CGC Arg	<del>,</del> H	GCC		GCC		GTC Val	GCC
	GGT	, <u>)</u>	•	CTC		GCC		GCT Ala	260	GTC	310 GAC ASP

FIGURE 6 1/5

						•						
	CAG Gln	450	CGG Arg		GCT		AAG Lys	GTC Val		ATC Ile	069	CTG
400	GGC Gly	7.	GAC ASP		AAG Lys		GAT Asp	590 CTA ACT Leu Thr	640	AAG Lys	w	GCG Ala
	GCC		AAC Asn		AAG Lys	540	ATT Ile	59( CTA 1 Leu		CGG Arg		TCT
	TTC	440	AAG Lys	490	66C G1y	u,	AAG Lys	66C 61y		CAC	089	GGG Gly
390	AGG Arg	44	GGC Gly		GCC Ala		TCC	GGT Gly	630	$_{\rm GGT}^{\rm GGT}$	99	ATG Met
1.7	ACC Thr		GAC		GTC Val	530	CTC	580 ATG Met	w w	AAA Lys		AAC Asn
	CCC		ATC Ile	480	ATT Ile	53	TCC	GGT Gly		GAG Glu		ACA Thr
380	TTC	430	TAC	7	TGC		CAA Gln	ACC Thr	620	ATC Ile	670	ATT
38	AAA Lys		66C G1y		TAC Tyr		66C G1y	570 GGA Gly	9	CTC		GCC Ala
	TCC		ACG Thr	470	CGC	520	GCC Ala	GTT Val		AAT Asn		TAT
	GCT	420	GCG Ala	47	CTC		CTC	CTA Leu		CAG Gln	* 099	CCA Pro
370	GAC	7.	AAC Asn		TGC		GAT ASP	560 GGA GTG Gly Val	610	GTT Val	w	ATT
	TTC Phe		TTC		GAT ASP	510	GCC	56 GGA G1Y		GGG		rrc Phe
	CGC Arg	410	GGC Gly	460	GAC Asp	u,	GAC Asp	GCC		GAC	20	TTT Phe
360	GAC	41	CGT Arg		CTC		GAA Glu	AGG Arg	009	TCT Ser	9	CCG
m	ATC Ile		ATC Ile		CGG Arg	200	CTC	550 GAG Glu	9	TTC Phe		TCC

FIGURE 6 2/5

	ACT		ATC	GCG		TCT	930	GAC		GAG Glu		TAT Tyr	
	TCA		CAT His	830 G GCT u Ala	880	TTA	on	AAG Lys		ATG Met		GAA Glu	
	ATT Ile	780	GCC AAT Ala Asn	83 GAG Glu		GCT Ala		GAT ASp		GTT Val	1020	GCA Ala	
730	TCG		GCC	ACT		AGG Arg	920	TGG Trp	970	TTG	10	ATT Ile	
	TAT		GCC	GGA Gly	870	TGC	9	CCG		GTA Val		ATT Ile	
	AAC Asn	770	GCT Ala	820 GGA G1y	ω	GCC Ala		AGG Arg		GGA Gly	0	CCG	
720	CCA	7.7	TAT Tyr	GCT		GTT Val		GCC TCA Ala Ser	096	GCT Ala	1010	GCG	v
	GGC		$ extsf{T}$	ATT	860	TTC	910		O.	GGG G1y		GGA Gly	TOTTO
	ATG Met		TGC	810 ATG Met	86	GGA Gly		ACT Thr		GAA Glu		CGG	<u>ن</u> 1
710	CTG Leu	760	TAC	CTG Leu		GGA Gly		CCT CAG Pro Gln	950	ATG GGT Met Gly	1000	AAA	
71	GGT Gly		AAC Asn	GAC Asp		TTA	006	CCT	9	ATG Met		ATG Met	
	TTG		TCC	)0 GCT Ala	850	GGT Gly	Oi	GAT		GTG Val		GCA ATG	
	GA'T Asp	750	ACT	800 GAG GC Glu A]		ATT Ile		GAT Asp		TTT Phe	066	CAT His	
700	ATC Ile		GCT	GGT Gly		CCA	0.0	AAT Asn	940	GGC Gly	01	GAG Glu	
	GCC		TGT Cys	CGA Arg	840	ATT Ile	89	AGG Arg		GAT Asp		TTG	
	CTT	740	GCA	790 CGC Arg	w	GTC Val		CAA		CGT Arg	086	AGC	

FIGURE 4/5

·											
AGG Arg		GAT Asp	1170	ACT		GTT Val		ATC Ile	ATT Ile		AAT Asn
1070 GAT CCA ASP Pro	1120	GAA Glu	11	GCG Ala		AAA GTT Lys Val		TCA ATG Ser Met	1310 GCA ACC Ala Thr	1360	CAA TTT Gln Phe
	•	CTC		CAT His		ATT AAG Ile Lys	1260	TCA	1310 GCA A		
ACT		AGT	20	GCT Ala	1210		ਜ	AAG Lys	ATC Ile		AAT Asn
ATG Met	1110	AGC	1160	AAT Asn	-	GCC		ACT	GCC	1350	ATT
1060 CAT His	H	GAG Glu		ATA Ile		AAT Asn	0.0	GCA	1300 GAA Glu	H	AGC
TAT Tyr		ATT Ile		TAC	1200	GAG ATA Glu Ile	1250	AAT	CTT		CCC
GCT	00	TGC Cys	1150	AAT Asn	7	GAG Glu		ATC Ile	GGT Gly	0 #	CAT His
)50 GAT ASP	1100	TCG Ser	-	GTC Val		CTT GCC Leu Ala		AAA Lys	1290 TCA GGA ( Ser Gly (	1340	CTT
1050 TGT GAT GCT Cys Asp Ala		TCC		GAG Glu	0.0	GAT CTT GCC Asp Leu Ala	1240	ACC AAG GAA ATC AAA ATC Thr Lys.Glu Ile Lys Ile	12 TCA Ser		TGG Trp
AAC Asn		GTC Val	1140	GAA Glu	1190	GAT	-	GAA Glu	GCA		GGC Gly
1040 GCA GTC Ala Val	1090	GGT	11	CCT		GGG		AAG Lys	1280 CTT GGA Leu Gly	1330	ACC ACC Thr Thr
1040 GCA G Ala Va	-	CTT Leu		TCA		CTT GCT Leu Ala	1230	ACC Thr	128 CTT Leu	-	ACC Thr
GGT Gly		GGG Gly	0	GTC Val	1180		12	AAC Asn	TGT Cys		ATA Ile
GGA G1y	1080	GAT Asp	1130	GGG G1y	-	ACT Thr		AAG Lys	CAC His	1320	GGA Gly
1030 TTG Leu	10	GCT		GCC		TCT	1220	TTC Phe	1270 GGA Gly	13	AAG Lys

SUBSTITUTE SHEET (RULE 26)

1410	s cag caa s Gln Gln		A GGG CAC	1510	ACTTGGTTCA	1570	TAAATGCCTT	1630	AGCCATTTAG	1690	СТСТВАТТТА	1750	GTTATTTAAG		CT
1400	AAC AAA AAG Asn Lys Lys	1450	GGA TTT GGA Gly Phe Gly	1500	TGA ATTCT	1560	AGCAATTTTT	1620	GTCCTTTGAT AGTTCCTCGA	1680	TAAATCTAGT	1740		1800	ATCCAGCTTA
1390	ACT GTT GCC Thr Val Ala	1440	AAT TCT TTT Asn Ser Phe	1490	TTC AAG CCA Phe Lys Pro	1550	AAATGCACAC CAGTTGCTGA GATAGGGCTT CAACTTGCAG AGCAATTTTT TAAATGCCTT	1610		1670	ATTCCCATTT TAAATCTAGT	1730	TGTATTAGAA AGACCAATGA AAGATTTTGT GTCATGTTTG TGTTGTCAAT	1790	ATAAAGCAAA AAAAAAAAA AAGGGCGGCC GCTCTAGAGG
	GAC TTC AAC ASP Phe Asn	1430	GCT ATC TCG Ala Ile Ser	1480	TTC TCA GCT Phe Ser Ala	1540	GATAGGGCTT	1600	CGTAATACCG GAATAGGTCG	1660	ATCGAAGATG	1720	AAGATTTTGT	1780	AAGGGCGGCC
1380	TCG GTG Ser Val		AAC GTC Asn Val	1470	GTG GCA Val Ala	1530	CAGTTGCTGA	1590		1650	TACTGTAATA	1710	AGACCAATGA	1770	AAAAAAAA
1370	CCC GAG CCA Pro Glu Pro	1420	CAT GAA GTG His Glu Val	1460	AAC TCG GTT Asn Ser Val	1520	AAATGCACAC	1580	GTCGGAAGAG	1640	GATGATGTTT	1700	TGTATTAGAA	1760	ATAAAGCAAA

FIGURE 6 5/5

Sequence Range: 1 to 2369

	GTACGCCTGC AGGTACCGGT CCGGAATTCC CGGGTCGACC CACGCGTCCG CATAAAAGAG	120	AGAGAGAGG ATCCATCGAA TGCGGCCACC CTCCTTTCAT CTTCGATTCA TTACCATACC	180	GCCTTTTCCG GGTCTTTCAT CCCAAAGGGT ATCCTTTTCT	TCC Ser>	. 280	TCT Ser>	330	CCT Pro>		CTA Leu>
	CATA		TTAC		ATCC	230 TCT Ser	7	ATG		TCT		CCA Pro
20	SCG (	110	rca ,	170	3GT	GCC		TGC	•	TCC	370	GCC
	CGTC	, ,	GAT	• •	AAAGC	GCC		GCC	320	ATC Ile	ω Γ	TGC
	CACC		CTTC		2000	CCT Pro	270	GCC	(*)	TCC ATC TCC Ser Ile Ser		CAA
40	ACC	100	CAT	160	CAT	220 CCTCCA ATG CCT Met Pro		CTT Leu		CCT		TCC
	GTCG		CTT		CTTI	CCA		CTC	0	CCG	360	CTC
	CGC		CTC		GGT		260	TGG	310	CTT Leu		ATT Ile
30	TTCC	90	CACC	150	TCCG	210 GTTC	7	ACG		CCT Pro		CGG ATT CTC TCC Arg Ile Leu Ser
	GGAA		CGGC		CTTT	210 CAGTCAGTTC		TGT Cys		GAC	350	CGC Arg
20	F CC	80	A TG		ပ္ပ		0	CTC	300	TCC	m	CGC
7	9922	∞	TCGA	140	TTTC	200 AGGGT	250	CCT		CCC		TCC
	GGTA		TCCA		ATTCCGCTGA TCCATTTTCC	200 CTCAAAGGGT		TCC		CAC	0	CTC
10	GC A	70	, 55°	130	GA 1			GCT	290	TTC Phe	340	CGA CGC CTC Arg Arg Leu
	GCCI		AGAG	П	CGCI	190 ATCCTATCTT	240	CTC	77	ACC.TCC Thr Ser		CGA Arg
	GTAC		AGAG		ATTC	ATCC		CTG		ACC		CGC Arg

FIGURE 7

*\( \frac{1}{2} \)* 

	GTC Val>	TCC Ser>	0	CGG Arg>	570	CTG Leu>		CAG Gln>		CAT His>	ATA Ile>
	CTC	470 ACA Thr	520	CAC His		GCT Ala		AAA Lys		GGC G1y	710 GGC G1y
420	ACC Thr	TAT TYF	•	AGG Arg		GTG Val	610	ATC Ile	* 099	CTA	AGT
	CAT His	TAC		CGC Arg	260	gcc Ala	61	AGT		CCT	ACG
	TTC	460 T GAC s Asp	510	ACC	u)	ATG Met		CCA		ACT	)0 GGA G1y
410	AGT	46 CAT His		ACC Thr		GCA Ala		AAG Lys	650	GTG Val	700 GAT GGA ASP Gly
4	TCC	TGC Cys		ATT CGC Ile Arg	0	GAG Glu	<b>600</b>	AAG Lys	W	GTG Val	CTT Leu
	GGA Gly	CCC	200	ATT Ile	52	AGG Arg		AAG Lys		GGT Gly	CTG
400	CGC Arg	450 GAG Glu	u)	CCC		TCC		ACA Thr	640	ATG Met	690 AAT Asn
40	CTC	TTC		AGA Arg		CCT Pro	290	ACC Thr	9	GGA Gly	AAT Asn
	GCC	TGC Cys	490	TCC	540 *	TCC	u,	GTT Val		ACT Thr	TAC
	TCC	440 GCC Ala	24	GGA Gly		GCT		GAA Glu		GTG Val	680 TTC Phe
390	TCC	CTC Leu		TTC		CGA Arg	580	CAG Gln	630	GTT Val	GTT Val
	GCT	TAC		TTG	530	AAT Asn	28	GAA Glu		GTA Val	GAT Asp
	TCT	TCT Ser	480	TCC	u,	CTC		CCT		CGA Arg	670 GAC CCT ASP Pro
380	CCT	430 ACC TCT Thr Sei		GCA		AGG Arg		CAA Gln	620	CGG Arg	670 GAC CO ASP PI

FIGURE 7

	•											
160	GCT Ala>	810	CTC Leu>		AAG Lys>		CTA Leu>	ATG Met>	00	AAG Lys>	1050	GCT Ala>
76	ATT Ile		AAG Lys		GGC Gly		GAG Glu	950 GGA Gly	1000	AAG Lys		TCA
	AGA Arg		CCG Pro	850	GCT	900	AAA Lys	GGT		TAT Tyr		GGA Gly
	ACG Thr	800	GCC Ala	8	ACC Thr		ATG Met	ATG Met		TCA	1040	ATG Met
750	CCT	~	GTG Val		CTG		GTG Val	940 TCA GCA	066	ATT Ile	1(	AAT
	TTT Phe		TGG		ATG Met	890	GAT Asp	94 TCA Ser		AGG Arg		ACA Thr
	CAA Gln	790	GGT Gly	840	TAC	~	GAA Glu	GGC Gly		CTA	30	GCT ACC Ala Thr
740	TGT GCT Cys Ala	7.	GAT		CTA		ACC	ATT Ile	086	GCC	1030	GCT
•			ACA Thr		ATG Met	880	ATC Ile	930 CTC Leu	0.	GAA		TTC
	GAT Asp		TCC	830	TTC	88	GGA Gly	GTT Val		ATT Ile		CCT
730	TTT Phe	780	TTC	~	AAG Lys		$_{\rm G1y}^{\rm GGT}$	GGA Gly	970	GCC Ala	1020	GTA
7.	ACC		TCT Ser		GAC		GAT Asp	920 TGC Cys	9	gat Asp	<b>v-</b> 1	TGT
	GAG Glu		AAG Lys	820	ATG Met	870	ACA Thr	AAA Lys		AAT Asn		$ ext{rr}$
	ATA Ile	170	ATC Ile	8	AGG Arg		TTA	aga Arg		TTC	1010	CCC
720	GAG Glu		GAG Glu		AAG Lys		GCA	LO AAA Lys	960	GTA Val	10	AAT Asn
	AGC		GGA Gly		TCT	860	AAA Lys	910 GAT AAI ASP LY		AAG Lys		ATG Met

FIGURE 7

FIGURE 7

										•		
	TCT Ser>		CAT His>	GCG Ala>	1240	TTG Leu>	1290	AGT Ser>		CTA Leu>		GAA Glu>
	ATA Ile		AAC	1190 A GAT E ASP	12	GCT		GAC		CTA		GCA Ala
0	TCG	1140	GCG	11 TCA Ser		CGA Arg		AGA CCA TGG Arg Pro Trp	30	CŢA	1380	ATT TAC Ile Tyr
1090	TAC	1-1	GCT	GGC Gly		TGC	1280	CCA	1330	GTG Val		ATT Ile
	AAC Asn		AAT Asn	30 GGG G1y	1230	GCA Ala	12			GGA Gly		ACT
	CCC Pro	1130	ATG Met	1180 TGC G( Cys G		GTT Val		TCA		GCT Ala	1370	GCG Ala
1080	GGG G1y	11	ATA Ile	CTT Leu		TTT Phe	0,	GCT	1320	GGA Gly	13	GGT Gly
П	ATG Met		TGT	ATG Met	1220	GGT Gly	1270	AAA Lys		GAA Glu		aga Arg
	TGG	0	${ m TTT}$	1170 GAT GTG ATG ASP Val Met	12	GGA Gly		ACT		GGG G1y	20	AAA Lys
1070	GGA Gly	1120	AAC Asn			ATG Met		GAC CCT Asp Pro	1310	ATG Met	1360	AAG
1(	TTG		AGT	GCA Ala	07	$_{\rm GLY}^{\rm GGT}$	1260	gac Asp	H	GTT Val		GCA
	GAC		ACG	1160 3C GAA 3Y Glu	1210	ATT Ile	•	TCC		TTT Phe		CAT His
0	ATG	1110	GCA	1160 GGC GAA Gly Glu		CCT		AAT Asn	0.0	GGA Gly	1350	GAG Glu
1060	GCA	Ε-1	$ ext{TGT}$	aga Arg		ATA Ile	1250	aga Arg	1300	GAT Asp	•	TTG
	CTT Leu		GCT	50 ATC Ile	1200	ATC Ile	12	CAG Gln		CGT		GAG Glu
	ATG Met	1100	ACT Thr	1150 ATA AT Ile I	П	GTA Val		TCC		AAT Asn	1340	GAG Glu

*								•				
gae ccr Glu Pro>	1480	TTG GCT Leu Ala>	1530	CAT GCC His Ala>		ATC CAC Ile His>		TCA ATG Ser Met>	1670 GTT TCA GTA Val Ser Val>	1720	TTG GAA Leu Glu>	
ACC Thr		GCT Ala		GCC	0	CTT Leu	1620	AAA Lys	16 GTT Val		AAT Asn	
GCC TAC CAC ATG ACC GAG Ala Tyr His Met Thr Glu		GAG AAG Glu Lys	1520	AAT Asn	1570	GCT CTT Ala Leu	-	AAT TCA ACC AAA TCA Asn Ser Thr Lys Ser	GCA Ala		ATT Ile	
CAC	1470	GAG Glu	15	ATA Ile		CAA Gln		TCA	1660 GTG GAA Val Glu	1710	CCG AAT Pro Asn	
TAC	(~1	ATA Ile		TAC		TAC	1610	AAT Asn	1660 GTG G2 Val G1	•••	CCG	
GCC Ala		TGC	0,	AAT Asn	1560	GAG Glu	1(	GTT Val	GGT Gly		CAT	
SAT	1460	CTC	1510	GTA Val	<b>(-1</b>	AAA Lys		AAA Lys	GGT Gly	1700	ATC Ile	
TGC	14	ATT Ile		GAC		ATC	00	TTA	1650 GCC	Ä	TGG	
ACT Thr		GTG		GAA Glu	1550	GAT Asp	1600	GAG Glu	GCA Ala		$\frac{GGG}{Gly}$	
rrc Phe	0.0	GGA Gly	1500	AGG	Η.	GGA Gly		aga Arg	GGA Gly	90	ACT Thr	
GGG AGT G	1450	GCT	П	TCT		GCT		AAC	1640 CTT CTC Leu Leu	1690	AGG	
14 GGG Gly		GGA		GTC Val	0,	CCG	1590	CAA	16 CTT Leu		ATA Ile	
3GT 31Y	•	GAT Asp	1490	GGA Gly	1540	ACT Thr	,,	GGC Gly	CAC		GCA Ala	
CTA	1440	CAC CCT His Pro	14	TCA		TCC		TTC Phe	30 GGT Gly	1680	cAG Gln	
1390 TTT CTA ( Phe Leu (	П	CAC His		CAG Gln		ACA Thr	1580	TGT Cys	1630 ATT GGT ( Ile Gly 1	•••	GTT Val	

FIGURE 7 5/7

1770	AAG AAG Lys Lys>		TTT GGT Phe Gly>	1870	GTTTCCGTGT	1930	GTTGGTAGCT	1990	GAACCATGAC	2050	GTAGAGCAAT	2110	GTTGTACTTT	2170	CACGTAGTAA
1760	GTG GGT CCT Val Gly Pro	1810	TCA TTT GGG Ser Phe Gly	1860	ATC TAG GAC Ile ***>	1920		1980	GCTTTAGTCG	2040	AGAATTGTTG	2100	TTTTTCTCTG AAATCTCCCT CCTTGCAATA	2160	TTAACTCGGG
1750	AAA TTG CTC Lys Leu Leu	1800	TTG TCT AAT Leu Ser Asn	1850	GCC CCT TAC Ala Pro Tyr	1910	AGTTTTGAGG	1970	TGTGTCCGGA	2030	ACTCCTTGCT	2090	AAATCTCCCT	2150	AACAAAGCTG
	SAT ACA Asp Thr	1790	GTC GGT Val Gly	1840	CTC TTC Leu Phe	1900	rcaaagctga	1960	CCATGAGTTT	2020	САСТТGАТАТ	2080	TTTTTCTCTG	2140	TGAAGAAGAG
1740	GAA GGC GTG GIU Glu Gly Val	1780	AAC GTT AAG Asn Val Lys	1830	TCG TCC ATA	1890	ACTCAACATA TCAAAGCTGA AGTTTTGAGG ACTCCAGCAT	1950	CTAGACATGC (	2010	CTCATGGCGA	2070	TCATATTTT '	2130	CGAGCTTTTC ATCGAGTCAG TGAAGAAGAG AACAAAGCTG
1730	AAC CCA GAT Asn Pro Asp	17	GAG AGA CTG Glu Arg Leu	1820	GGG CAC AAC Gly His Asn	1880	GTGGAATTCT	1940	CCTTACGTCT	2000	GGATTGAGTA	2060	ATTCATTATC	2120	CGAGCTTTTC

FIGURE 7 6/7

7	
FIGURE	717

2290 ATGTATGTTT 2350 AAAAAAAAA	2280 * TAATTGGGGR 2340 * AAAAAAAAA	2270 TTCTCATTGA 2330 AAAAAAAAA	2240 2250 2260 2270 2280 2290 2290 **  AACTAGAAGA CTGGTTTAGA TTGGTTTGTT TTCTCATTGA TAATTGGGGR ATGTATGTTT	2250 CTGGTTTAGA 2310 AAAAAAAAA	2240 AGAAGA 2300 AATAAA
2290	2280	2270	2260	2250	2240
aaatttgtaa	TGTGGTTTTA	ATCACCGTTT	CCATITIGCCC TITGITITIGC ICICIATITIC AICACCGITI IGIGGITITIA AAAITIGIAA	TTTGTTTTGC	၁၁၁၅
2230	2220	2210	2200	2190	2180

2360 AGGGCGCCG CTCTAGAGG

Sequence Range: 1 to 2374

									_		٠.	<b>-</b> -	<i>r</i> >	O +	<b>~</b>
*	CACACCAAAC	120	ACAGACAGAC	180	TCTTCGATTC	240	TCCCAAAGGG	300	CCTGCCGCCT	360	TCTACCTCCT	420	CICICCCGC	480	CGCGGATCC
50	GACGCCAACC	110	AGACAGACAG	170	CCTCCTTTCA	230	GGGTCTTTCA	290	CCCTCCAATG	350	CGCCTGCATG	410	TCGCCGACGC	470	CTCCGCCCTC
40	ACGCGTCCGC	100	CATTGGCAGC	160	ATGCGGCCAC	220	CGCCTTTTCC GGGTCTTTCA TCCCAAAGGG	280	TCAGTCAGTT	340	GGCTCCTTGC	400	CCGCCTTCCA TCTCCTCTCC TCGCCGACGC CTCTCCCGCC	460	CTTCTGCTTC
30	GGGTCGACCC	06	AGACGGACGC	150	GATCCATCGA	210		270	TATCCTATCT TCTCAAAGGG TCAGTCAGTT	330	CTCTGTACGT	390		450	CICCCAAIGC GCCCCACTAC CITCIGCIIC CICCGCCCIC CGCGGAICCA
20	CGGAATTCCC GGGTCGACCC ACGCGTCCGC GACGCCAACC CACACCAAAC	80	TTCCTCAGCT TCTCTTCTCA AGACGGACGC CATTGGCAGC AGACAGACAG ACAGACAGAC	140	CCATAAAAGA GAGAGAGG GATCCATCGA ATGCGGCCAC CCTCCTTTCA TCTTCGATTC	200	CATTCCGCTG ATCCATTTTC	260		320	CTTCCCTGCT CGCTTCCCCT CTCTGTACGT	380	CGACCCTCTT	440	
10	-A-CNTGGTC	70	TTCCTCAGCT	130	CCATAAAAGA	190	ATTACCATAC	250	TATCCTTTTC	310	CTTCCCTGCT	370	TCCACCCTC	430	GCCGGATTCT

FIGURE 8

CAGCAATGGG	CTCATTGGCT	ATGCGGAGTT	AAAGAGCTAG ATAAAAGAAA		AGATGTGATG
1020	1010	1000	066	980	970
GAATCACCGA	ACAGATGGTG	GAAAGCATTA	CTGCTGGCAA	TACATGCTGA	GTTCATGCTA
096	950	940	930	920	910
CTCTCTAAGA GGATGGACAA	CTCTCTAAGA	ATGGTTGGGT GGCCCCGAAG	ATGGTTGGGT	TTCTCCACAG	GATCAAGTCT
006	890	880	870	860	850
TGCTCAATTT CCTACGAGAA TTGCTGGAGA	CCTACGAGAA		GAGATAGAGA CCTTTGATTG		TGGCATAAGC
840	830	. 820	810	800	790
ATGGAACGAG	AATCTGCTTG	GIGACTCCTC TAGGCCAIGA ACCTGATGIT TITCTACAAI AATCTGCTTG	ACCTGATGTT	TAGGCCATGA	GTGACTCCTC
780	770	760	750	740	730
AATGGGTGTG	TTGTGACTGG	CGGCGAGTAG	TATCAAACAG	ACCACAAAGA AGAAGCCAAG TATCAAACAG	ACCACAAAGA
720	710	700	069	089	049
ACAGGAAGTT	TGCAACCTGA	GGAGGCAATG GCCGTGGCTC		CCCTTCCAGG	ATCGAGCTTC
* 099	650	640	630	. 620	610
CGGAGGCTCA	CCGCAGGCAC	TTCGCACCAC	TCCAGACCCA	CTTGTTCGGA	CATCCGCATC
*	590	580	570	260	550
GACTACTATA	GCCCTGCCAT	CCTGCTTCGA GCCCTGCCAT GACTACTATA	TCTTACCTCG	CCTCGTCACC	GTTTCCATAC
540	530	520	510	200	490

FIGURE 8 2/5

TGAA	7.40	* * * * * * * * * * * * * * * * * * *	ACTT	1200	GTAT	1260	BGCTC	1320	rccca	1380	TTTGT	1440	AGAGG	1500	ATGAC
	AGAAGA		CAATGGACTT		ACTTT		999909		CTTTG		ATGGA		AGAAA		ACCAC
KE KOMUU -	AT'I'I'CATALA	1130	GCTATGCTTG	1190	GCAACGAGTA	1250	GTGATGCTTT	1310	GCATGCCGAG CTTTGTCCCA	1370	AGTAATCGTG	1430	GAGCATGCAA	1490	TGCGATGCCT
	TGGAATGAAG GTATTCAATG ATGCCATTGA AGCCCTAAGG ATTTCATATA AGAAGALGAA	1120	CTACCACAAA TATGGGATCA GCTATGCTTG	1180	GGGCCCAACT ACTCGATATC TACTGCTTGT GCAACGAGTA ACTTTTGTAT	1240	AATGAATGCT GCGAACCATA TAATCAGAGG CGAAGCAGAT GTGATGCTTT GCGGGGGCTC	1300	AGATGCGGTA ATCATACCTA TTGGTATGGG AGGTTTTGTT	1360	GAGAAATTCC GACCCTACTA AAGCTTCAAG ACCATGGGAC AGTAATCGTG ATGGATTTGT	1420	TAIGGGGGAA GGAGCIGGAG IGCIACIACI AGAGGAGITG GAGCAIGCAA AGAAAAGAGG	1480	TGCGACTATT TACGCAGAAT TTCTAGGTGG GAGTTTCACT TGCGATGCCT ACCACATGAC
	ATGCCA'I"I'GA	1110	CTACCACAAA	1170	ACTCGATATC	1230	TAATCAGAGG	1290	TTGGTATGGG	1350	AAGCTTCAAG	1410	TGCTACTACT	1470	TTCTAGGTGG
	GTATTCAATG	1100	GTACCTTTCG	1160	GGGCCCAACT	1220	GCGAACCATA	1280	ATCATACCTA	1340	GACCCTACTA	1400	GGAGCTGGAG	1460	TACGCAGAAT
	TGGAATGAAG	1090	TCCCTTTTGT	1150	GGGATGGATG	1210	AATGAATGCT	1270	AGATGCGGTA	1330	GAGAAATTCC	1390	TATGGGGGAA	1450	TGCGACTATT

FIGURE 8 3/5

TTTGTGTCCG	GCCCATGAGT	GGACTCCAGC ATGITGGIAG CICCTIACGI CICTAGACAI GCCCAIGAGI ITIGIGICCG	CTCCTTACGT	ATGTTGGTAG	GGACTCCAGC
2040	2030	2020	2010	2000	1990
GAAGTTTTGA	TATCAAAGCT	GTGTGGAATT CTACTCAACA TATCAAAGCT GAAGTTTTGA	GTGTGGAATT	TTACATCTAG GACGTTTCGT	TTACATCTAG
1980	1970	1960	1950	1940	1930
TCTTCGCCCC	TCGTCCATAC	TCTAATICAT ITGGGITTIGG IGGGCACAAC ICGICCATAC ICTICGCCCC	TTGGGTTTGG		GGTCGGTTTG
1920	1910	1900	1890	1880	1870
TGAACGTTAA	AAGGAGAGAC	AGATGAAGGC GTGGATACAA AATTGCTCGT GGGTCCTAAG AAGGAGAGAC TGAACGTTAA	AATTGCTCGT	GTGGATACAA	AGATGAAGGC
1860	1850	1840	1830	1820	1810
TGGAAAACCC	AATATTAATT	GATCCATCCG	CAGGCAATAA GGACTGGGTG		TTCAGTAGTT
1800	1790	1780	1770	1760	1750
TGGAAGCAGT	GCCGGTGGTG	TCTCGGAGCA GCCGGTGGTG TGGAAGCAGT	AAATCAATGA TTGGTCACCT	AAATCAATGA	TAATTCAACC
1740	1730	1720	1710	1700	1690
AGTTAAAAGT	CAAAACAGAG	AGATATCAAA GAGTACCAAG CTCTTATCCA CTGTTTCGGC CAAAACAGAG AGTTAAAAGT	CTCTTATCCA	GAGTACCAAG	AGATATCAAA
1680	1670	1660	1650	1640	1630
CTCCGGCTGG	GCCACATCCA CTCCGGCTGG	AAATGCCCAT		AGGAGTCTCT AGGGAAGACG TAAATTACAT	AGGAGTCTCT
1620	1610	1600	1590	1580	1570
TGGCTCAGTC	GAGAAGGCTT	CTGGAGTGAT TCTCTGCATA GAGAAGGCTT TGGCTCAGTC	CTGGAGTGAT	CGAGCCTCAC CCTGATGGAG	CGAGCCTCAC
1560	1550	1540	1530	1520	1510

FIGURE

		ATCC	2350 2360 2370 AAAAAAAAA AAGGGCGGCC GCTCTAGAGG	2360 AAGGGCGGCC	2350 AAAAAAAAA
TTTTCTCAAA	GATTGGTTTG	GACTGGTTTA	TITGIGGITI TAAAATITGI AAAACTAGAA GACTGGITTA GATIGGITIG ITTICICAAA	TAAAATTTGT	TTTGTGGTTT
2340	2330	2320	2310	2300	2290
TCATCACCGT	GCTCTCTATT	CCTTTGTTTT	TGTTAACTCG GGCACGTAGT AACCATTTGC CCTTTGTTTT GCTCTCTATT TCATCACCGT	GGCACGTAGT	TGTTAACTCG
2280	2270	2260	2250	2240	2230
AGAACAAAGC	AGTGAAGAAG	TCATCGAGTC	CTCCTTGCAA TAGTTGTACT TTCGAGCTTT TCATCGAGTC AGTGAAGAAG AGAACAAAGC	TAGTTGTACT	CTCCTTGCAA
2220	2210	2200	2190	2180	2170
TGAAATCTCC	TTTTTTCTC	TCTCATATTT	CTAGAATTGT TGGTAGAGCA ATATTCATTA TCTCATATTT TTTTTTTCTC TGAAATCTCC	TGGTAGAGCA	CTAGAATTGT
2160	2150	2140	2130	2120	2110
ATACTCCTTG	GACACTTGAT	TACTCATGGC	GAGCTTTAGT CGGAACCATG ACGGATTGAG TACTCATGGC GACACTTGAT ATACTCCTTG	CGGAACCATG	GAGCTTTAGT
2100	2090	2080	2070	2060	2050

FIGURE 8 5/5

Sequence Range: 1 to 1580

^		Α.Δ	_	. ^.		r. A		s. A	4 A		, <u>A</u>
GGG G1y>	100	TCG Ser>	150	GTG Val>		GAT Asp>		TCT Ser>	AAA Lys>	340	CGC Arg>
50 TCT Ser	1(	CAT		AGG Arg		GGT Gly		GGA Gly	290 GCT Ala	m	ATC CGC Ile Arg
GCA Ala		CAG Gln		AAA Lys	o	TTG	240	ATT Ile	CTT CEU	•	GGG
AAT Asn		ACT Thr	140	TCC	190	TCT		TTA	GAT Asp		ACG Thr
40 GCG Ala	90	GCA Ala	П	GTC Val		CAG Gln		AAA Lys	30 GAT Asp	330	CGA Arg
ATG Met		AGG Arg		TTT Phe		AGG	230	TGC	280 AAT GP ASn As		GTC Val
റ്ദേദ		AGA Arg	130	GAG	180	GAC	.,	GGA Gly	TCA		ACT
GC3	80	CTG	ਜ	TCG		TCT		AGA Arg	GTC	320	ATT Ile
30 CGTT		GCC		TCC		GAT Asp	220	AGT	270 CAA Gln		TGG
GTTI		CCT		TCT	170	CAG Gln	22	GTG Val	CTT Leu		GAA Glu
10 20 30 CCTGAATCGG ATTCAAGAGA GAGTTTCGTT GCTGGG	70	GTT Val	120	GGA Gly	-	GTT Val		CTT	GCT	310	AAT GAT Asn Asp
2 AGAG	(-	TCA		CGT Arg		GCC		CCG AGG Pro Arg	260 CCA Pro	, M	
TTC		TCT		TCT Ser	160	AGT	210		ATA Ile		ACC
10 3GG 2		GGT Gly	110	TCG	16	TGT Cys		TCG	GCT		GAC
AATC	÷	CTG Leu	177	TCA		TGC Cys		CGC	250 GGT TCT Gly Ser	300	GTC
CCTG		TTT Phe	•	ATT Ile		TTT Phe	200	TCT	25 GGT G1Y		ATT

FIGURE 9 1/5

390	TCA Ser>		GAT Asp>		GGC G1y>	TTG Leu>	280	GTC Val>	630	GTG Val>		GGA Gly>	
	GCA Ala		AAT Asn		TTC Phe	530 AAT CCT Asn.Pro	28	TTA Leu		CTA		CGG Arg	
	TTA Leu	0	GCA	480	CTT			GGT Gly		ATT Ile	019	GAT Asp	
380	AAT Asn	430	GAC		GAC	AAG Lys		TTG	620	AAT Asn	9	ACC Thr	
m	ACA	ı	GTA Val		GAG Glu	520 TGC AAA AA Cys Lys Ly	570	GTG Val	v	AAC Asn		TGG Trp	
	CTT Leu		CAG Gln	470	CCT	52 TGC Cys		TTT Phe		TTT Phe		GAC	
0	AGT	420	GCA Ala	4,	ACC Thr	960 G1y		GGA Gly	610	$_{\rm GGT}^{\rm GGT}$	<b>660</b>	GTT Val	
370	GAT Asp		ATG		TCT	CTT	260	AGT	61	666 61y		TAT	
	AAA Lys		GAG Glu	460	ACT Thr	510 GCA Ala	u,	TGC		$_{\rm GGT}^{\rm GGT}$		CGG	
	$_{\rm GLY}^{\rm GGT}$	410	CTA	46	TGT	AAA Lys		GCA		ATT AGA Ile Arg	650	TCT	
360	TCA	4	GCT		ATG Met	TCG	550	GCT	009	ATT Ile	v	CTT	
	CTC		AAA Lys		TTG	00 ATA Ile	5.5	ACC Thr		CAC His		TCT	
	GTT Val	400	AGG	450	GTT Val	5 CAG Gln		ATT Ile		TGC	640	GAT Asp	
350	AGG Arg	40	GCA		ATG Met	CCT		GAC Asp	590	GCT Ala	9	GCT	
(•)	CGA Arg		GCA Ala		GAT Asp	490 AGT GCT Ser Ala	540	TAC TYr		GCT Ala		GGT Gly	
	AAC		GAG Glu	440	GTG Val	490 AGT G		TCT		TCA		ATT	

FIGURE 9 2/5

	TCA Ser>	GAT Asp>	820	GTT Val>	870	AGG Arg>		CGC Arg>		AAG Lys>	GCA Ala>
	CAG Gln	770 AGC Ser	8	gaa Glu		CCA		TTC		GGA Gly	010 CAG Gln
720	GTG Val	7 CAT His		GAT Asp		CCA	910	GTA Val	960	CTT Leu	1( CAT His
	GTG Val	TTG Leu		gaa Glu	860	TTT Phe	91	GAG Glu		GCA Ala	CTT Leu
	GTA Val	760 TTT GAT '	810	aaa Lys	ω	GAT Asp		AAA Lys		TCA	1000 TTG CTG Leu Leu
710	GCT	76 TTT Phe		ATC Ile		aga Arg		$_{\rm G1y}^{\rm GGT}$	950	GAA Glu	100 TTG Leu
	GGA Gly	GCT		GCA Ala	850	ATC Ile	900	AAC Asn	O1	ATC Ile	TGG
	GCT	TTT Phe	800	GCT	8	TCC		ATG		TCA	GAC Asp
700	GCT	750 CTC Leu	w	AAA Lys		GGG		CAA	940	CAG Gln	990 ATC Ile
7(	'GAT Asp	GGG G1y		CTA		AAT Asn	890	ATC	6	CCT	AAC Asn
	GGA Gly	GAT Asp	790	CAT His	840	CAT	w	TGC		GTG Val	TCC
	TTT Phe	740 GAA Glu	7.5	AGG Arg		GGA Gly		TCT		TCT	980 GGA G1y
069	CTC	7 GAG Glu		CAA		CTG	880	TAC	930	CGC	AAT Asn
	ATT	GCT		GGG Gly	830	GCC	88	TCA		TGC Cys	CTT Leu
	TGT	730 3T GAT 7S ASP	780	GAT Asp	w	AAA Lys		TCT		GCT	970 C GGT a Gly
089	ACA Thr	73 TGT Cys		GGA G1y		GAT		CGT Arg	920	$ ext{rr}$	97 GCC Ala

FIGURE 9 3/5

1060	CCT CAA Pro Gln>	1110	GCG GCA Ala Ala>		GTG AAG Val Lys>		ACA TGG Thr Trp>	1260	CACTGCAGCT	1320	AAGAAGTCAG	1380	TCGTTCCCCT
1050	CTA GAG GTT Leu Glu Val	1100	AAC ACT AGT Asn Thr Ser	1150	AGT GGA AAT Ser Gly Asn	1200	GCC GGA CTC Ala Gly Leu	1250	GGA TAA GACTGAA GCCGAGCCAG Gly ***>	1310	CCANAAAAAG	1370	TTGCCCTTTT
	ACA CGT Thr Arg	1090	TAC GGG Tyr Gly	1140	GTG AGG Val Arg	1190	TTT GGC Phe Gly	1240	A GACTGAA *>	1300	GCTTCCATGA	1360	TTCATCACA
1040	GCA GTA GCA Ala Val Ala	Н	TTG GCA AAT Leu Ala Asn	1130	GAC GAA GCT Asp Glu Ala	1180	ACC GCA GGA Thr Ala Gly	1230	TGG GGA TA Trp Gly **	1290	ACGAAATTTT G	1350	GACACGAT C
1030	ATT GAT Ile Asp	1080	ATC TCA AAC Ile Ser Asn		TTG GCA CTA Leu Ala Leu	70	GTG ATT GCA Val Ile Ala	1220	GCT ATT ATC AGG Ala Ile Ile Arg	1280	CCGATGTTTC AC	1340	CAAGCAAC AC
1020	AAT CAG AGG ATC Asn Gln Arg Ile	1070	GAA CGA ATT A Glu Arg Ile I	1120	TCC ATT CCC T Ser Ile Pro L	1160 1170	CCG GGT CAC G Pro Gly His V	1210	GGT TCT GCT A Gly Ser Ala I	1270	TCCTCTCAAA CC	1330	TCTTTTATGG AGCAAGCAAC ACGACACGAT CTTCATCACA TTGCCCTTTT TCGTTCCCCT
						11							

FIGURE 9

1440	TTGTCCCCAA	1500	CGGGACATTG	1560	AAAAAAAA	
1430	ATAGTTTCTT	1490	CATTTTGTCT	1550	AAAAAAAAA	
1420	TACAATACCC	1480	GCTTTTACTT	1540	TTTGCTAAAA	
1410	TTGCTGACAA	1470	TAATTGTTCA	1530	ATGTTTATAT	
1400	TITCCATTAG TITGATGATT TIGCTGACAA TACAATACCC ATAGTITCTT TIGICCCAA	1460	TAAGTTATTT GTTTCTTGTT TAATTGTTCA GCTTTTACTT CATTTTGTCT CGGGACATTG	1520	GAGATGACAG CATAAACATC ATGTTTATAT TTTGCTAAAA AAAAAAAAA AAAAAAAA	1580 AAAAAAAAA
1390	TTTCCATTAG	1450	TAAGTTATTT	1510	GAGATGACAG	1570 1580 AAAAAAAA AAAAAAAA

FIGURE 9 5/5

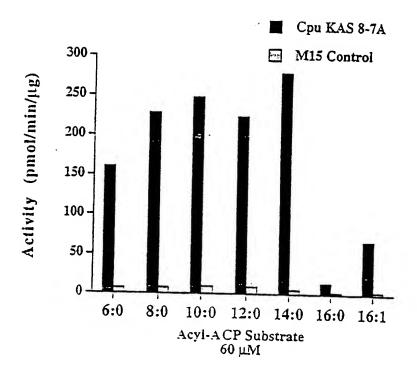


FIGURE 10

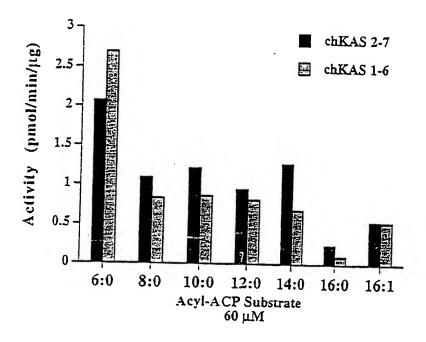


FIGURE 11

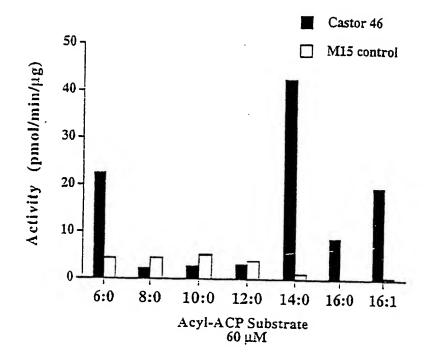
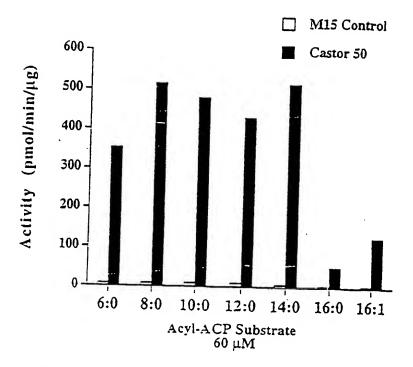


FIGURE 12



E328013-28

FIGURE 13

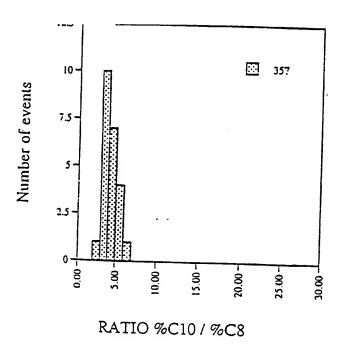


FIGURE 15

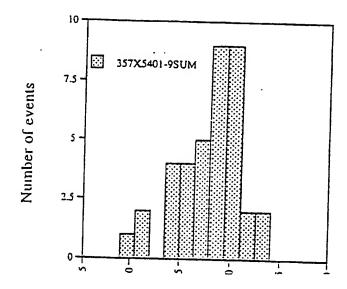
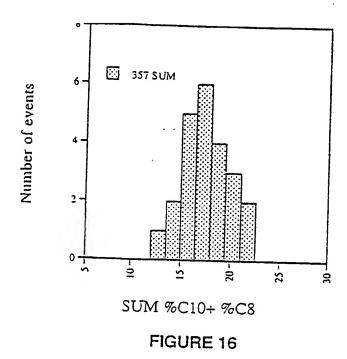


FIGURE 15 2/2



SUBSTITUTE SHEET (RULE 26)

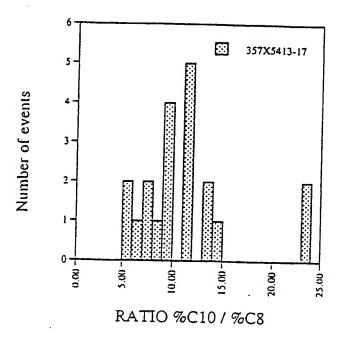


FIGURE 17 1/2

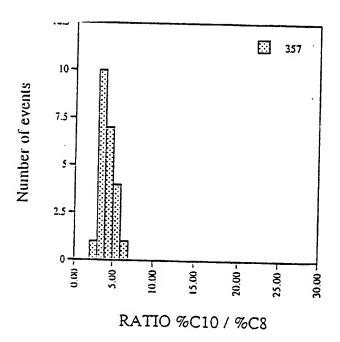


FIGURE 17

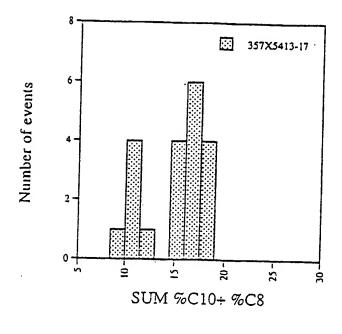


FIGURE 18 1/2

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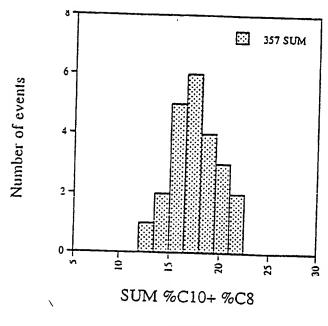
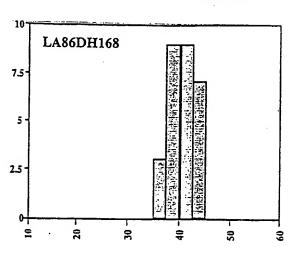


FIGURE 18





### 12:0 levels (w%)

FIGURE 19 1/3



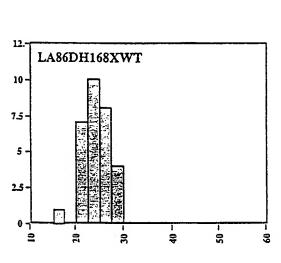
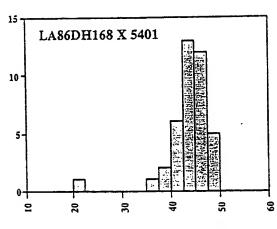


FIGURE 19 3/3 SUBSTITUTE SHEET (RULE 26)



12:0 levels (w%)

FIGURE 19 2/3

SUBSTITUTE SHEET (RULE 26)

WO 98/46776

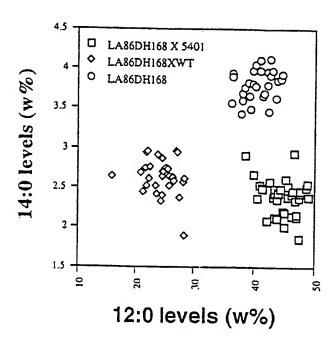
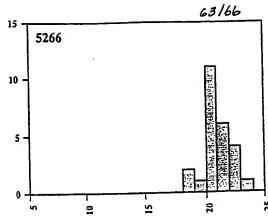


FIGURE 20

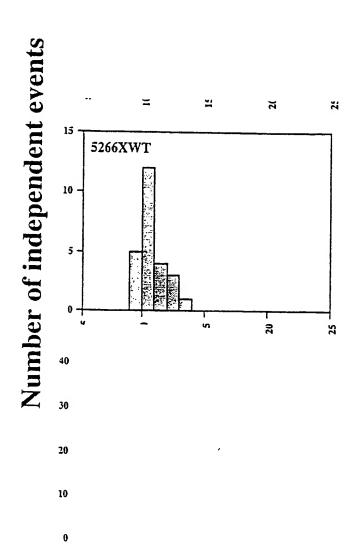




18:0 levels (w%)

FIGURE -21

1/3



18:0 levels (w%)

<u>FIGURE 21</u> 2/3

Number of independent events

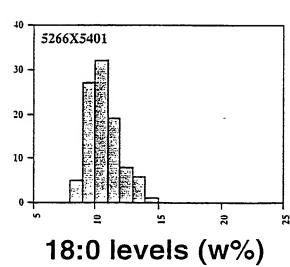


FIGURE 21 3/3

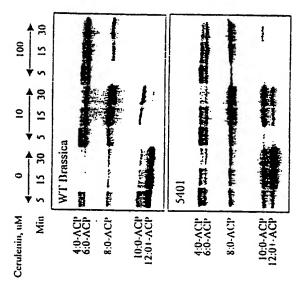


FIGURE 22

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